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(54) Title: OB FUSION PROTEIN COMPOSITIONS AND METHODS

(57) Abstract

The present invention relates to Fc-OB fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to a genetic or chemical fusion protein comprising the Fc immunoglobulin region, derivative or analog fused to the N-terminal portion of the OB protein, derivative or analog.

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OB FUSION PROTEIN COMPOSITIONS AND METHODS

Field of the Invention

The present invention relates to Fc-OB fusion protein compositions and methods for preparation and use thereof.

Background

Although the molecular basis for obesity is largely unknown, the identification of the "OB gene" and protein encoded ("OB protein" or "leptin") has shed some light on mechanisms the body uses to regulate body fat deposition. See, PCT publication, WO 96/05309

(12/22/96), Friedman et al.; Zhang et al., Nature 372: 425-432 (1994); see also, the Correction at Nature 374: 479 (1995). The OB protein is active in vivo in both

ob/ob mutant mice (mice obese due to a defect in the production of the OB gene product) as well as in normal, wild type mice. The biological activity manifests itself in, among other things, weight loss. See generally, Barrinaga, "Obese" Protein Slims Mice, Science 269: 475-456 (1995). The OB protein, derivatives and use thereof as modulators for the control of weight and adiposity of animals, including mammals and humans, has been disclosed in greater detail in PCT publication WO 96/05309 (12/22/96), hereby incorporated by reference, including figures.

The other biological effects of OB protein are not well characterized. It is known, for instance, that in ob/ob mutant mice, administration of OB protein results in a decrease in serum insulin levels, and serum glucose levels. It is also known that administration of OB protein results in a decrease in body fat. This was observed in both ob/ob mutant mice, as well as non-obese normal mice. Pelleymounter et al., Science 269: 540-543

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(1995); Halaas et al., Science 269: 543-546 (1995). See also, Campfield et al., Science 269: 546-549 (1995) (Peripheral and central administration of microgram doses of OB protein reduced food intake and body weight of ob/ob and diet-induced obese mice but not in db/db obese mice.) In none of these reports have toxicity's been observed, even at the highest doses.

Despite the promise of clinical application of the OB protein, the mode of action of the OB protein <u>in</u> <u>vivo</u> is not clearly elucidated. Information on the OB receptor, shows high affinity binding of the OB protein detected in the rat hypothalamus, which indicates OB receptor location. Stephens et al., Nature <u>377</u>: 530-532. The <u>db/db</u> mouse displays the identical phenotype as the <u>ob/ob</u> mouse, <u>i.e.</u>, extreme obesity and Type II diabetes; this phenotype is thought to be due to a defective OB receptor, particularly since <u>db/db</u> mice fail to respond to OB protein administration. <u>See</u> Stephens et al., <u>supra</u>.

With the advances in recombinant DNA technologies, the availability of recombinant proteins for therapeutic use has engendered advances in protein formulation and chemical modification. One goal of such modification is protein protection and decreased degradation. Fusion proteins and chemical attachment may effectively block a proteolytic enzyme from physical contact with the protein backbone itself, and thus prevent degradation. Additional advantages include, under certain circumstances, increasing the stability, circulation time, and the biological activity of the therapeutic protein. A review article describing protein modification and fusion proteins is Francis,

Focus on Growth Factors $\underline{3}$:4-10 (May 1992) (published by Mediscript, Mountview Court, Friern Barnet Lane, London N20, OLD, UK).

One such modification is the use of the Fc
region of immunoglobulins. Antibodies comprise two
functionally independent parts, a variable domain known
as "Fab", which binds antigen, and a constant domain,
known as "Fc" which provides the link to effector
functions such as complement or phagocytic cells. The
Fc portion of an immunoglobulin has a long plasma halflife, whereas the Fab is short-lived. Capon, et al.,
Nature 337: 525-531 (1989).

Therapeutic protein products have been constructed using the Fc domain to provide longer halflife or to incorporate functions such as Fc receptor 15 binding, protein A binding, complement fixation and placental transfer which all reside in the Fc proteins of immunoglobulins. Id. For example, the Fc region of an IgG1 antibody has been fused to the N-terminal end of CD30-L, a molecule which binds CD30 receptors expressed 20 on Hodgkin's Disease tumor cells, anaplastic lymphoma cells, T-cell leukemia cells and other malignant cell types. See, U.S. Patent No. 5,480,981. IL-10, an antiinflammatory and antirejection agent has been fused to 25 murine Fcγ2a in order to increase the cytokine's short circulating half-life. Zheng, X. et al., The Journal of Immunology, $\underline{154}$: 5590-5600 (1995). Studies have also evaluated the use of tumor necrosis factor receptor linked with the Fc protein of human IgG1 to treat patients with septic shock. Fisher, C. et al., N. Engl. 30 J. Med., 334: 1697-1702 (1996); Van Zee, K. et al., The Journal of Immunology, <u>156</u>: 2221-2230 (1996). Fc has also been fused with CD4 receptor to produce a therapeutic protein for treatment of AIDS. See, Capon et al., Nature, 337:525-531 (1989). In addition, the N-terminus of interleukin 2 has also been fused to the

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Fc portion of IgG1 or IgG3 to overcome the short half life of interleukin 2 and its systemic toxicity. See, Harvill et al., Immunotechnology, 1: 95-105 (1995).

Due to the identification of the OB protein as a promising therapeutic protein, there exists a need to 5 develop OB analog compositions for clinical application in conjunction with or in place of OB protein administration. Such development would include OB analog compositions where protein formulations and chemical modifications achieve decreased protein 10 degradation, increased stability and circulation time. The present invention provides such compositions.

Summary of the Invention

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The present invention relates to Fc-OB fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to a genetic fusion protein comprising the Fc region or analogs of immunoglobulins fused to the N-terminal portion of the OB protein or analogs. The Fc-OB fusion protein is capable of dimerizing via the cysteine residues of the Fc region. Unexpectedly, genetic fusion modification with Fc at the N-terminus of the OB protein demonstrates advantages in stability, clearance rate and decreased degradation which are not seen in OB protein or with fusion of Fc to the C-terminus of the OB protein. Surprisingly and importantly, the N-terminus modification provides unexpected protein protection from degradation, increases circulation time and stability, when compared to the OB protein or Fc modification to the OB protein C-terminus. Such unexpected advantages from the Fc modification to OB protein would be advantageous to OB protein consumers, in that these changes contribute to lower doses required or less frequent dosing. Thus, as

described below in more detail, the present invention

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has a number of aspects relating to the genetic modification of proteins via fusion of the Fc region to the OB protein (or analogs thereof), as well as, specific modifications, preparations and methods of use thereof.

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Accordingly, in one aspect, the present invention provides a Fc-OB fusion protein wherein Fc is genetically fused to the N-terminus of the OB protein (or analogs thereof). In addition, the Fc portion may also be linked to the N-terminus of the OB protein (or analogs thereof) via peptide or chemical linkers as known in the art. As noted above and described in more detail below, the Fc-OB fusion protein has unexpected protections from degradation and increased circulation time and stability when compared to the OB protein or C-terminus OB-Fc fusion proteins. Additional aspects of the present invention, therefore, include not only Fc-OB fusion protein compositions, but also DNA sequences encoding such proteins, related vectors and host cells containing such vectors, both useful for producing fusion proteins of the present invention.

In a second aspect, the present invention provides for preparing the Fc-OB fusion protein. Such methods include recombinant DNA techniques for preparation of recombinant proteins. Furthermore, such aspects include methods of fermentation and purification as well.

In another aspect, the present invention provides methods for treating excess weight in an individual or animals, including modulation of and/or fat deposition by the administration of Fc-OB fusion proteins. Due to the Fc-OB fusion protein characteristics, methods are contemplated which reduce the amount and/or frequency of dosage of OB protein by using Fc-OB weight reducing agent.

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In yet another aspect, the present invention provides for therapies for the treatment of co-morbidities associated with excess fat, such as diabetes, dys- or hyperlipidemias, arterial sclerosis, arterial plaque, the reduction or prevention of gall stones formation, stoke, and also an increase in insulin sensitivity and/or an increase in lean tissue mass.

In another aspect, the present invention also provides for related pharmaceutical compositions of the Fc-OB proteins, analogs and derivatives thereof, for use 10 in the above therapies.

Brief Description of the Drawings

FIGURE 1 Recombinant murine metOB (double 15 stranded) DNA (SEQ. ID. NOs.: 1 and 2) and amino acid sequence (SEQ. ID. NO. 3).

FIGURE 2 Recombinant human metOB analog (double stranded) DNA (SEQ. ID. NOs.: 4 and 5) and amino acid sequence (SEQ. ID. NO. 6).

20 FIGURE 3 (A-C) Recombinant human metFc-OB (double stranded) DNA (SEQ. ID. NOs.: 7 and 8) and amino acid sequence (SEQ. ID. NO. 9).

FIGURE 4 (A-C) Recombinant human metFc-OB variant (double stranded) DNA (SEQ. ID. NOs.: 10 and 11) and amino acid sequence (SEQ. ID. NO. 12).

FIGURE 5 (A-C) Recombinant human metFc-OB variant (double stranded) DNA (SEQ. ID. NOs.: 13 and 14) and amino acid sequence (SEQ. ID. NO. 15).

FIGURE 6 (A-C) Recombinant human metFc-OB variant (double stranded) DNA (SEQ. ID. NOs.: 16 and 17) 30 and amino acid sequence (SEQ. ID. NO. 18).

<u>Detailed Description</u>

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The present invention relates to Fc-OB fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the

present invention relates to the genetic or chemical fusion of the Fc region of immunoglobulins to the N-terminal portion of the OB protein. Unexpectedly, fusion of Fc at the N-terminus of the OB protein

5 demonstrates advantages which are not seen in OB protein or with fusion of Fc at the C-terminus of the OB protein. Surprisingly, the N-terminally modified Fc-OB protein provides unexpected protein protection from degradation, increased circulation time and increased stability. Accordingly, the Fc-OB fusion protein, and analogs or derivatives thereof, as well as, related methods of use and preparation, are described in more detail below.

15 <u>Compositions</u>

The Fc sequence of the recombinant human Fc-OB sequence set forth in SEQ. ID. NO. 9 (See Figure 3) may be selected from the human immunoglobulin IgG-1 heavy chain, see Ellison, J.W. et al., Nucleic Acids Res. 10: 4071-4079 (1982), or any other Fc sequence known in the art (e.g. other IgG classes including but not limited to IgG-2, IgG-3 and IgG-4, or other immunoglobulins). Variant, analogs or derivatives of the Fc portion may be constructed by, for example, making various substitutions of residues or sequences.

Cysteine residues can be deleted or replaced with other amino acids to prevent formation of disulfide crosslinks of the Fc sequences. In particular amino acid at position 5 of SEQ. ID. NO. 9 is a cysteine residue. The recombinant Fc-OB sequence of SEQ. ID. NO. 9 is a 378 amino acid Fc-OB protein (not counting the methionine residue). The first amino acid sequence for the recombinant Fc-OB protein of Figure 3 is referred to as +1 with the methionine at the -1 position.

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One may remove the cysteine residue at position 5 or substitute it with one or more amino acids. An alanine residue may be substituted for the cysteine residue at position 6 giving the variant amino acid sequence of Figure 4 (SEQ. ID. NO. 12). The recombinant Fc-OB protein of Figure 4 is a 378 amino acid Fc-OB protein (not counting the methionine residue). The first amino acid sequence for the recombinant Fc-OB protein of Figure 4 is referred to as +1 with the methionine at the -1 position.

Likewise, the cysteine at position 5 of SEQ.

ID. NO. 9 could be substituted with a serine or other amino acid residue or deleted. A variant or analog may also be prepared by deletion of amino acids at positions 1, 2, 3, 4 and 5 as with the variant in SEQ. ID. NO. 15 (See Figure 5). Substitutions at these positions can also be made and are with in the scope of this invention. The recombinant Fc-OB protein of Figure 5 is a 373 amino acid Fc-OB protein (not counting the methionine residue). The first amino acid sequence for the recombinant Fc-OB protein of Figure 5 is referred to as +1 with the methionine at the -1 position.

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Modifications may also be made to introduce four amino acid substitutions to ablate the Fc receptor binding site and the complement (Clq) binding site. These variant modifications from SEQ. ID. NO. 15 would include leucine at position 15 substituted with glutamate, glutamate at position 98 substituted with alanine, and lysines at positions 100 and 102 substituted with alanines (see Figure 6 and SEQ. ID. NO. 18). The recombinant Fc-OB protein of Figure 6 is a 373 amino acid Fc-OB protein (not counting the methionine residue). The first amino acid sequence for the recombinant Fc-OB protein of Figure 6 is referred to as +1 with the methionine at the -1 position.

Likewise, one or more tyrosine residues can be replaced by phenyalanine residues as well. In addition, other variant amino acid insertions, deletions and/or substitutions are also contemplated and are within the scope of the present invention. Furthermore, alterations may be in the form of altered amino acids, such as peptidomimetics or D-amino acids. The Fc protein may be also linked to the OB proteins of the Fc-OB protein by "linker" moieties whether chemical or amino acids of varying lengths. Such chemical linkers 10 are well known in the art. Amino acid linker sequences can include but are not limited to:

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	(a)	ala, ala, ala;
	(b)	ala, ala, ala;
15	(c)	ala, ala, ala, ala;
	(d)	gly, gly;
	(e)	gly, gly, gly;
	(f)	gly, gly, gly, gly;
	(g)	gly, gly, gly, gly, gly, gly;
20	(h)	gly-pro-gly;
	(i)	gly, gly, pro, gly, gly; and
	(j)	any combination of subparts (a)
thro	ough (i).	

The OB portion of the Fc-OB fusion protein may be selected from the recombinant murine set forth in 25 SEQ. ID. NO. 3 (See Figure 1), or the recombinant human protein as set forth in Zhang et al., Nature, supra, (herein incorporated by reference) or those lacking a glutaminyl residue at position 28. (See Zhang et al, Nature, supra, at page 428.) One may also use the 30 recombinant human OB protein analog as set forth in SEQ. ID. NO. 6 (See Figure 2), which contains: (1) an arginine in place of lysine at position 35; and (2) a leucine in place of isoleucine at position 74. (A shorthand abbreviation for this analog is the recombinant human $R->L^{35}$, $I->L^{74}$). The amino acid

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sequences for the recombinant human and recombinant murine proteins or analogs with or without the fused Fc portion at the N-terminus of the OB protein are set forth below with a methionyl residue at the -1 position; however, as with any of the present OB proteins and analogs, the methionyl residue may be absent.

The murine protein is substantially homologous to the human protein, particularly as a mature protein, and, further, particularly at the N-terminus. One may prepare an analog of the recombinant human protein by altering (such as substituting amino acid residues), in the recombinant human sequence, the amino acids which diverge from the murine sequence. Because the recombinant human protein has biological activity in mice, such an analog would likely be active in humans. For example, using a human protein having a lysine at residue 35 and an isoleucine at residue 74 according to the numbering of SEQ. ID. NO. 6, wherein the first amino acid is valine, and the amino acid at position 146 is cysteine, one may substitute with another amino acid one or more of the amino acids at positions 32, 35, 50, 64, 68, 71, 74, 77, 89, 97, 100, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145. One may select the amino acid at the corresponding position of the murine protein, (SEQ. ID. NO. 3), or another amino acid.

One may further prepare "consensus" molecules based on the rat OB protein sequence. Murakami et al., Biochem. Biophys. Res. Comm. 209: 944-952 (1995) herein incorporated by reference. Rat OB protein differs from human OB protein at the following positions (using the numbering of SEQ. ID. NO. 6): 4, 32, 33, 35, 50, 68, 71, 74, 77, 78, 89, 97, 100, 101, 102, 105, 106, 107, 108, 111, 118, 136, 138 and 145. One may substitute with another amino acid one or more of the amino acids at these divergent positions. The positions in bold print are those in which the murine OB protein as well as the

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rat OB protein are divergent from the human OB protein, and thus, are particularly suitable for alteration. At one or more of a positions, one may substitute an amino acid from the corresponding rat OB protein, or another amino acid.

The positions from both rat and murine OB protein which diverge from the mature human OB protein are: 4, 32, 33, 35, 50, 64, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145. An OB protein according to SEQ. ID. NO. 6 10 having one or more of the above amino acids replaced with another amino acid, such as the amino acid found in the corresponding rat or murine sequence, may also be effective.

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- 15 In addition, the amino acids found in rhesus monkey OB protein which diverge from the mature human OB protein are (with identities noted in parentheses in one letter amino acid abbreviation): 8 (S), 35 (R), 48(V), 53(Q), 60(I), 66(I), 67(N), 68((L), 89(L), 100(L), 20 108(E), 112 (D), and 118 (L). Since the recombinant human OB protein is active in cynomolgus monkeys, a human OB protein according to SEQ. ID. NO. 6 (with lysine at position 35 and isoleucine at position 74) having one or more of the rhesus monkey divergent amino acids replaced with another amino acid, such as the amino acids in parentheses, may be effective. It should be noted that certain rhesus divergent amino acids are also those found in the above murine species (positions 35, 68, 89, 100 and 112). Thus, one may prepare a murine/rhesus/human consensus molecule having (using the 30 numbering of SEQ. ID. NO. 6 having a lysine at position 35 and an isoleucine at position 74) having one or more of the amino acids at positions replaced by another
- amino acid: 4, 8, 32, 33, <u>35</u>, 48, 50, 53, 60, 64, 66, 67, <u>68</u>, 71, 74, 77, 78, <u>89</u>, 97, <u>100</u>, 102, 105, 106, 107, 108, 111, <u>112</u>, 118, 136, 138, 142, and 145.

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Other analogs may be prepared by deleting a part of the protein amino acid sequence. For example, the mature protein lacks a leader sequence (-22 to -1). One may prepare the following truncated forms of human OB protein molecules (using the numbering of SEQ. ID. NO. 6):

- (a) amino acids 98-146
- (b) amino acids 1-32
- (c) amino acids 40-116
- 10 (d) amino acids 1-99 and (connected to) 112-146
 - (e) amino acids 1-99 and (connected to) 112-146 having one or more of amino acids 100-111 placed between amino acids 99 and 112.
- In addition, the truncated forms may also have 15 altered one or more of the amino acids which are divergent (in the rat, murine, or rhesus OB protein) from human OB protein. Furthermore, any alterations may be in the form of altered amino acids, such as 20 peptidomimetics or D-amino acids.

Therefore, the present invention encompasses a Fc-OB fusion protein wherein the OB protein is selected from:

- the amino acid sequence 1-146 as set 25 forth in SEQ. ID. NO. 3 (below) or SEQ. ID. NO. 6;
 - the amino acid sequence 1-146 as set forth in SEQ. ID. NO. 6 having a lysine residue at position 35 and an isoleucine residue at position 74;
- (c) the amino acid sequence of subpart (b) 30 having a different amino acid substituted in one or more of the following positions (using the numbering according to SEQ. ID. NO. 6 and retaining the same numbering even in the absence of a glutaminyl residue at position 28): 4, 32, 33, 35, 50, 64, 68, 71, 74, 77,
- 35 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145;

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(d) the amino acid sequence of subparts (a),(b) or (c) optionally lacking a glutaminyl residue at position 28;

- (e) the amino acid sequence of subparts (a),(b), (c), or (d) having a methionyl residue at theN-terminus;
 - (f) a truncated OB protein analog selected from among: (using the numbering of SEQ. ID. NO. 6):
 - (i) amino acids 98-146

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- (ii) amino acids 1-32
- (iii) amino acids 40-116
- (iv) amino acids 1-99 and 112-146
- (v) amino acids 1-99 and 112-146 having one or more of amino acids 100-111 placed between amino acids 99 and 112; and,
- (vi) the truncated OB analog of subpart(i) having one or more of amino acids 100, 102,105, 106, 107, 108, 111, 118, 136, 138, 142, and145 substituted with another amino acid;

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- (vii) the truncated analog of subpart
 (ii) having one or more of amino acids 4, 8 and 32
 substituted with another amino acid;
- (viii) the truncated analog of subpart (iii) having one or more of amino acids 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111 and 112 replaced with another amino acid;
 - (vix) the truncated analog of subpart (iv) having one or more of amino acids 4, 8, 32, 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 112, 118, 136, 138, 142, and 145 replaced with another amino acid; and
 - (x) the truncated analog of subpart (v) having one or more of amino acids 4, 32, 33, 35,

50, 64, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 118, 136, 138, 142, and 145 replaced with another amino acid;

- (xi) the truncated analog of any of subparts (i)-(x) having an N-terminal methionyl residue; and
- the OB protein or analog derivative of any of subparts (a) through (f) comprised of a chemical moiety connected to the protein moiety;
- 10 a derivative of subpart (g) wherein said chemical moiety is a water soluble polymer moiety;
 - a derivative of subpart (h) wherein said water soluble polymer moiety is polyethylene glycol;
 - (j) a derivative of subpart (h) wherein said water soluble polymer moiety is a polyaminoacid moiety;
 - a derivative of subpart (h) through (j) wherein said moiety is attached at solely the N-terminus of said protein moiety; and
- (1) an OB protein, analog or derivative of any of subparts (a) through (k) in a pharmaceutically 20 acceptable carrier.

<u>Derivatives</u>

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The present Fc-OB fusion proteins (herein the term "protein" is used to include "peptide," Fc, OB or 25 analogs, such as those recited infra, unless otherwise indicated) are derivatized by the attachment of one or more chemical moieties to the Fc-OB fusion protein moiety. These chemically modified derivatives may be 30 further formulated for intraarterial, intraperitoneal, intramuscular subcutaneous, intravenous, oral, nasal, pulmonary, topical or other routes of administration as discussed below. Chemical modification of biologically active proteins has been found to provide additional advantages under certain circumstances, such as increasing the stability and circulation time of the

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therapeutic protein and decreasing immunogenicity. See, U.S. Patent No. 4,179,337, Davis et al., issued December 18, 1979. For a review, see Abuchowski et al., in Enzymes as Drugs. (J. S. Holcerberg and J. Roberts, eds. pp. 367-383 (1981)); Francis et al., supra.

The chemical moieties suitable for such derivatization may be selected from among various water soluble polymers. The polymer selected should be water soluble so that the protein to which it is attached does not precipitate in an aqueous environment, such as a 10 physiological environment. Preferably, for therapeutic use of the end-product preparation, the polymer will be pharmaceutically acceptable. One skilled in the art will be able to select the desired polymer based on such 15 considerations as whether the polymer/protein conjugate will be used therapeutically, and if so, the desired dosage, circulation time, resistance to proteolysis, and other considerations. For the present proteins and peptides, the effectiveness of the derivatization may be 20 ascertained by administering the derivative, in the desired form (i.e., by osmotic pump, or, more preferably, by injection or infusion, or, further formulated for oral, pulmonary or nasal delivery, for example), and observing biological effects as described herein.

The water soluble polymer may be selected from the group consisting of, for example, polyethylene glycol, copolymers of ethylene glycol/propylene glycol, carboxymethylcellulose, dextran, polyvinyl alcohol, 30 polyvinyl pyrolidone, poly-1, 3-dioxolane, poly-1,3,6-trioxane, ethylene/maleic anhydride copolymer, polyaminoacids (either homopolymers or random copolymers), and dextran or poly(n-vinyl pyrolidone)polyethylene glycol, propylene glycol homopolymers, polypropylene oxide/ethylene oxide co-polymers, polyoxyethylated polyols and polyvinyl

alcohol. Polyethylene glycol propionaldenhyde may have advantages in manufacturing due to its stability in water. Also, succinate and styrene may also be used.

The OB or Fc proteins used to formulate the 5 Fc-OB fusion protein, may be prepared by attaching polyaminoacids or branch point amino acids to the Fc or OB protein (or analogs) moiety. For example, the polyaminoacid may be an additional carrier protein which, like the Fc fused to the OB protein or OB analog 10 of the present invention, serves to also increase the circulation half life of the protein in addition to the advantages achieved via the Fc-OB fusion protein above. For the present therapeutic or cosmetic purpose of the present invention, such polyaminoacids should be those which have or do not create neutralizing antigenic 15 response, or other adverse responses. Such polyaminoacids may be selected from the group consisting of serum album (such as human serum albumin), an additional antibody or portion thereof (e.g. the Fc 20 region), or other polyaminoacids, e.g. lysines. As indicated below, the location of attachment of the polyaminoacid may be at the N-terminus of the Fc-OB protein moiety, or C-terminus, or other places in between, and also may be connected by a chemical 25 "linker" moiety to the Fc-OB protein.

The polymer may be of any molecular weight, and may be branched or unbranched. For polyethylene glycol, the preferred molecular weight is between about 2 kDa and about 100 kDa (the term "about" indicating that in preparations of polyethylene glycol, some molecules will weigh more, some less, than the stated molecular weight) for ease in handling and manufacturing. Other sizes may be used, depending on the desired therapeutic profile (e.g., the duration of sustained release desired, the effects, if any on biological activity, the ease in handling, the degree or

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lack of antigenicity and other known effects of the polyethylene glycol to a therapeutic protein or analog).

The number of polymer molecules so attached may vary, and one skilled in the art will be able to ascertain the effect on function. One may mono-derivatize, or may provide for a di-, tri-, tetraor some combination of derivatization, with the same or different chemical moieties (e.g., polymers, such as different weights of polyethylene glycols). proportion of polymer molecules to protein (or peptide) 10 molecules will vary, as will their concentrations in the reaction mixture. In general, the optimum ratio (in terms of efficiency of reaction in that there is no excess unreacted protein or polymer) will be determined by factors such as the desired degree of derivatization (e.g., mono, di-, tri-, etc.), the molecular weight of the polymer selected, whether the polymer is branched or unbranched, and the reaction conditions.

The chemical moieties should be attached to 20 the protein with consideration of effects on functional or antigenic domains of the protein. There are a number of attachment methods available to those skilled in the art. <u>E.g.</u>, EP 0 401 384 herein incorporated by reference (coupling PEG to G-CSF), see also Malik et al., Exp. 25 Hematol. <u>20</u>: 1028-1035 (1992) (reporting pegylation of GM-CSF using tresyl chloride). For example, polyethylene glycol may be covalently bound through amino acid residues via a reactive group, such as, a free amino or carboxyl group. Reactive groups are those 30 to which an activated polyethylene glycol molecule may be bound. The amino acid residues having a free amino group may include lysine residues and the N-terminal amino acid residue. Those having a free carboxyl group may include aspartic acid residues, glutamic acid residues, and the C-terminal amino acid residue. Sulfhydryl groups may also be used as a reactive group

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for attaching the polyethylene glycol molecule(s). Preferred for therapeutic purposes is attachment at an amino group, such as attachment at the N-terminus or lysine group. Attachment at residues important for receptor binding should be avoided if receptor binding is desired.

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One may specifically desire N-terminally chemically modified Fc-OB fusion protein. Using polyethylene glycol as an illustration of the present 10 compositions, one may select from a variety of polyethylene glycol molecules (by molecular weight, branching, etc.), the proportion of polyethylene glycol molecules to protein (or peptide) molecules in the reaction mix, the type of pegylation reaction to be 15 performed, and the method of obtaining the selected N-terminally pegylated protein. The method of obtaining the N-terminally pegylated preparation (i.e., separating this moiety from other monopegylated moieties if necessary) may be by purification of the N-terminally pegylated material from a population of pegylated 20 protein molecules. Selective N-terminal chemical modification may be accomplished by reductive alkylation which exploits differential reactivity of different types of primary amino groups (lysine versus the N-terminal) available for derivatization in a particular 25 protein. Under the appropriate reaction conditions, substantially selective derivatization of the protein at the N-terminus with a carbonyl group containing polymer is achieved. For example, one may selectively N-terminally pegylate the protein by performing the 30 reaction at a pH which allows one to take advantage of the pK_a differences between the ϵ -amino group of the lysine residues and that of the α -amino group of the N-terminal residue of the protein. By such selective derivatization, attachment of a water soluble polymer to a protein is controlled: the conjugation with the

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polymer takes place predominantly at the N-terminus of the protein and no significant modification of other reactive groups, such as the lysine side chain amino groups, occurs. Using reductive alkylation, the water soluble polymer may be of the type described above, and should have a single reactive aldehyde for coupling to the protein. Polyethylene glycol propionaldehyde, containing a single reactive aldehyde, may be used.

An N-terminally monopegylated derivative is

10 preferred for ease in production of a therapeutic.

N-terminal pegylation ensures a homogenous product as characterization of the product is simplified relative to di-, tri- or other multi-pegylated products. The use of the above reductive alkylation process for

15 preparation of an N-terminal product is preferred for ease in commercial manufacturing.

Complexes

The Fc-OB fusion protein, analog or derivative thereof may be administered complexed to a binding 20 composition. Such binding composition may have the effect of prolonging the circulation time even further than that achieved with the Fc-OB fusion protein, analog or derivative. Such composition may be a protein (or synonymously, peptide). An example of a binding 25 protein is OB protein receptor or portion thereof, such as a soluble portion thereof. Other binding proteins may be ascertained by examining OB protein or Fc-OB protein in serum, or by empirically screening for the presence of binding. Binding proteins used will 30 typically not interfere with the ability of OB protein, Fc-OB fusion proteins, or analogs or derivatives thereof, to bind to endogenous OB protein receptor and/or effect signal transduction.

Pharmaceutical Compositions

The present invention also provides methods of using pharmaceutical compositions of the Fc-OB fusion proteins and derivatives. Such pharmaceutical compositions may be for administration for injection, or for oral, pulmonary, nasal, transdermal or other forms of administration. In general, comprehended by the invention are pharmaceutical compositions comprising effective amounts of protein or derivative products of 10 the invention together with pharmaceutically acceptable diluents, preservatives, solubilizers, emulsifiers, adjuvants and/or carriers. Such compositions include diluents of various buffer content (e.g., Tris-HCl, acetate, phosphate), pH and ionic strength; additives such as detergents and solubilizing agents (e.g., Tween 15 80, Polysorbate 80), anti-oxidants (e.g., ascorbic acid, sodium metabisulfite), preservatives (e.g., Thimersol, benzyl alcohol) and bulking substances (e.g., lactose, mannitol); incorporation of the material into particulate preparations of polymeric compounds such as 20 polylactic acid, polyglycolic acid, etc. or into liposomes. Hylauronic acid may also be used, and this may have the effect of promoting sustained duration in the circulation. Such compositions may influence the 25 physical state, stability, rate of in vivo release, and rate of in vivo clearance of the present proteins and derivatives. <u>See</u>, <u>e.g.</u>, Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712 which are herein incorporated by reference. The compositions may be prepared in liquid 30 form, or may be in dried powder, such as lyophilized Implantable sustained release formulations are form. also contemplated, as are transdermal formulations.

Contemplated for use herein are oral solid

dosage forms, which are described generally in

Remington's Pharmaceutical Sciences, 18th Ed. 1990 (Mack

Publishing Co. Easton PA 18042) at Chapter 89, which is herein incorporated by reference. Solid dosage forms include tablets, capsules, pills, troches or lozenges, cachets or pellets. Also, liposomal or proteinoid encapsulation may be used to formulate the present 5 compositions (as, for example, proteinoid microspheres reported in U.S. Patent No. 4,925,673). Liposomal encapsulation may be used and the liposomes may be derivatized with various polymers (e.g., U.S. Patent No. 5,013,556). A description of possible solid dosage 10 forms for the therapeutic is given by Marshall, K. In: Modern Pharmaceutics Edited by G. S. Banker and C. T. Rhodes Chapter 10, 1979, herein incorporated by reference. In general, the formulation will include the Fc-OB fusion protein (or analog or derivative), and 15 inert ingredients which allow for protection against the stomach environment, and release of the biologically active material in the intestine.

Also specifically contemplated are oral dosage forms of the above derivatized proteins. Fc-OB fusion 20 protein may be chemically modified so that oral delivery of the derivative is efficacious. Generally, the chemical modification contemplated is the attachment of at least one moiety to the protein (or peptide) molecule itself, where said moiety permits (a) inhibition of 25 proteolysis; and (b) uptake into the blood stream from the stomach or intestine. Also desired is the increase in overall stability of the protein and increase in circulation time in the body. Examples of such moieties include: Polyethylene glycol, copolymers of ethylene 30 glycol and propylene glycol, carboxymethyl cellulose, dextran, polyvinyl alcohol, polyvinyl pyrrolidone and polyproline. Abuchowski and Davis, Soluble Polymer-Enzyme Adducts. In: "Enzymes as Drugs", Hocenberg and Roberts, eds., Wiley-Interscience, New York, NY, (1981), pp. 367-383; Newmark, et al., J. Appl.

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Biochem. $\underline{4}$: 185-189 (1982). Other polymers that could be used are poly-1,3-dioxolane and poly-1,3,6-tioxocane. Preferred for pharmaceutical usage, as indicated above, are polyethylene glycol moieties.

for the Fc-OB fusion protein, analog or derivative, the location of release may be the stomach, the small intestine (e.g., the duodenum, jejunum, or ileum), or the large intestine. One skilled in the art has available formulations which will not dissolve in the stomach, yet will release the material in the duodenum or elsewhere in the intestine. Preferably, the release will avoid the deleterious effects of the stomach environment, either by protection of the Fc-OB fusion protein, analog or derivative, or by release of the biologically active material beyond the stomach environment, such as in the intestine.

To ensure full gastric resistance a coating impermeable to at least pH 5.0 is essential. Examples of the more common inert ingredients that are used as enteric coatings are cellulose acetate trimellitate (CAT), hydroxypropylmethylcellulose phthalate (HPMCP), HPMCP 50, HPMCP 55, polyvinyl acetate phthalate (PVAP), Eudragit L30D, Aquateric, cellulose acetate phthalate (CAP), Eudragit L, Eudragit S, and Shellac. These coatings may be used as mixed films.

A coating or mixture of coatings can also be used on tablets, which are not intended for protection against the stomach. This can include sugar coatings, or coatings which make the tablet easier to swallow. Capsules may consist of a hard shell (such as gelatin) for delivery of dry therapeutic i.e. powder; for liquid forms, a soft gelatin shell may be used. The shell material of cachets could be thick starch or other edible paper. For pills, lozenges, molded tablets or tablet triturates, moist massing techniques can be used.

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The therapeutic can be included in the formulation as fine multiparticulates in the form of granules or pellets of particle size about 1 mm. The formulation of the material for capsule administration could also be as a powder, lightly compressed plugs or even as tablets. The therapeutic could be prepared by compression.

Colorants and flavoring agents may all be included. For example, the protein (or derivative) may be formulated (such as by liposome or microsphere encapsulation) and then further contained within an edible product, such as a refrigerated beverage containing colorants and flavoring agents.

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One may dilute or increase the volume of the

therapeutic with an inert material. These diluents

could include carbohydrates, especially mannitol,

α-lactose, anhydrous lactose, cellulose, sucrose,

modified dextrans and starch. Certain inorganic salts

may be also be used as fillers including calcium

triphosphate, magnesium carbonate and sodium chloride.

Some commercially available diluents are Fast-Flo,

Emdex, STA-Rx 1500, Emcompress and Avicell.

Disintegrants may be included in the formulation of the therapeutic into a solid dosage form. 25 Materials used as disintegrates include but are not limited to starch including the commercial disintegrant based on starch, Explotab. Sodium starch glycolate, Amberlite, sodium carboxymethylcellulose, ultramylopectin, sodium alginate, gelatin, orange peel, acid carboxymethyl cellulose, natural sponge and 30 bentonite may all be used. Another form of the disintegrants are the insoluble cationic exchange resins. Powdered gums may be used as disintegrants and as binders and these can include powdered gums such as agar, Karaya or tragacanth. Alginic acid and its sodium salt are also useful as disintegrants.

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Binders may be used to hold the therapeutic agent together to form a hard tablet and include materials from natural products such as acacia, tragacanth, starch and gelatin. Others include methyl cellulose (MC), ethyl cellulose (EC) and carboxymethyl cellulose (CMC). Polyvinyl pyrrolidone (PVP) and hydroxypropylmethyl cellulose (HPMC) could both be used in alcoholic solutions to granulate the therapeutic.

An antifrictional agent may be included in the
formulation of the therapeutic to prevent sticking
during the formulation process. Lubricants may be used
as a layer between the therapeutic and the die wall, and
these can include but are not limited to; stearic acid
including its magnesium and calcium salts,
polytetrafluoroethylene (PTFE), liquid paraffin,

polytetrafluoroethylene (PTFE), liquid paraffin, vegetable oils and waxes. Soluble lubricants may also be used such as sodium lauryl sulfate, magnesium lauryl sulfate, polyethylene glycol of various molecular weights, Carbowax 4000 and 6000.

Glidants that might improve the flow properties of the drug during formulation and to aid rearrangement during compression might be added. The glidants may include starch, talc, pyrogenic silica and hydrated silicoaluminate.

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To aid dissolution of the therapeutic into the aqueous environment a surfactant might be added as a wetting agent. Surfactants may include anionic detergents such as sodium lauryl sulfate, dioctyl sodium sulfosuccinate and dioctyl sodium sulfonate. Cationic detergents might be used and could include benzalkonium chloride or benzethomium chloride. The list of potential nonionic detergents that could be included in the formulation as surfactants are lauromacrogol 400, polyoxyl 40 stearate, polyoxyethylene hydrogenated castor oil 10, 50 and 60, glycerol monostearate,

polysorbate 40, 60, 65 and 80, sucrose fatty acid ester,

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methyl cellulose and carboxymethyl cellulose. These surfactants could be present in the formulation of the protein or derivative either alone or as a mixture in different ratios.

Additives which potentially enhance uptake of the protein (or derivative) are for instance the fatty acids oleic acid, linoleic acid and linolenic acid.

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Controlled release formulation may be desirable. The drug could be incorporated into an inert matrix which permits release by either diffusion or leaching mechanisms e.g., gums. Slowly degenerating matrices may also be incorporated into the formulation, e.g., alginates, polysaccahrides. Another form of a controlled release of this therapeutic is by a method based on the Oros therapeutic system (Alza Corp.), i.e., the drug is enclosed in a semipermeable membrane which allows water to enter and push drug out through a single small opening due to osmotic effects. Some enteric coatings also have a delayed release effect.

Other coatings may be used for the formulation. These include a variety of sugars which could be applied in a coating pan. The therapeutic agent could also be given in a film coated tablet and the materials used in this instance are divided into 2 groups. The first are the nonenteric materials and include methyl cellulose, ethyl cellulose, hydroxyethyl cellulose, methylhydroxy-ethyl cellulose, hydroxypropyl cellulose, hydroxypropyl-methyl cellulose, sodium carboxy-methyl cellulose, providone and the polyethylene glycols. The second group consists of the enteric materials that are commonly esters of phthalic acid.

A mix of materials might be used to provide the optimum film coating. Film coating may be carried out in a pan coater or in a fluidized bed or by compression coating.

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Also contemplated herein is pulmonary delivery of the present protein (or derivatives thereof). The protein (or derivative) is delivered to the lungs of a mammal while inhaling and traverses across the lung epithelial lining to the blood stream. (Other reports

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- epithelial lining to the blood stream. (Other reports of this include Adjei et al., Pharmaceutical Research 7: 565-569 (1990); Adjei et al., International Journal of Pharmaceutics 63: 135-144 (1990) (leuprolide acetate); Braquet et al., Journal of Cardiovascular
- Pharmacology 13 (suppl. 5): s.143-146 (1989) (endothelin-1); Hubbard et al., Annals of Internal Medicine 3: 206-212 (1989) (α1-antitrypsin); Smith et al., J. Clin. Invest. 84: 1145-1146
- (1989)(α-1-proteinase); Oswein et al., "Aerosolization
 of Proteins", Proceedings of Symposium on Respiratory
 Drug Delivery II, Keystone, Colorado, March, 1990
 (recombinant human growth hormone); Debs et al., The
 Journal of Immunology 140: 3482-3488 (1988) (interferon-γ
 and tumor necrosis factor α) and Platz et al., U.S.
- Patent No. 5,284,656 (granulocyte colony stimulating factor).

Contemplated for use in the practice of this invention are a wide range of mechanical devices designed for pulmonary delivery of therapeutic products, including but not limited to nebulizers, metered dose inhalers, and powder inhalers, all of which are familiar to those skilled in the art.

Some specific examples of commercially available devices suitable for the practice of this invention are the Ultravent nebulizer, manufactured by Mallinckrodt, Inc., St. Louis, Missouri; the Acorn II nebulizer, manufactured by Marquest Medical Products, Englewood, Colorado; the Ventolin metered dose inhaler, manufactured by Glaxo Inc., Research Triangle Park,

North Carolina; and the Spinhaler powder inhaler, manufactured by Fisons Corp., Bedford, Massachusetts.

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All such devices require the use of formulations suitable for the dispensing of protein (or analog or derivative). Typically, each formulation is specific to the type of device employed and may involve the use of an appropriate propellant material, in addition to diluents, adjuvants and/or carriers useful in therapy.

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The protein (or derivative) should most advantageously be prepared in particulate form with an average particle size of less than 10 μm (or microns), most preferably 0.5 to 5 μm , for most effective delivery to the distal lung.

Carriers include carbohydrates such as trehalose, mannitol, xylitol, sucrose, lactose, and sorbitol. Other ingredients for use in formulations may include DPPC, DOPE, DSPC and DOPC. Natural or synthetic surfactants may be used. Polyethylene glycol may be used (even apart from its use in derivatizing the protein or analog). Dextrans, such as cyclodextran, may be used. Bile salts and other related enhancers may be used. Cellulose and cellulose derivatives may be used. Amino acids may be used, such

Also, the use of liposomes, microcapsules or microspheres, inclusion complexes, or other types of carriers is contemplated.

as use in a buffer formulation.

romulations suitable for use with a nebulizer, either jet or ultrasonic, will typically comprise Fc-OB protein, analogs or derivatives thereof, dissolved in water at a concentration of about 0.1 to 25 mg of biologically active protein per mL of solution. The formulation may also include a buffer and a simple sugar (e.g., for protein stabilization and regulation of osmotic pressure). The nebulizer formulation may also contain a surfactant, to reduce or prevent surface

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induced aggregation of the protein caused by atomization of the solution in forming the aerosol.

Formulations for use with a metered-dose inhaler device will generally comprise a finely divided powder containing the protein (or derivative) suspended in a propellant with the aid of a surfactant. The propellant may be any conventional material employed for this purpose, such as a chlorofluorocarbon, a hydrochlorofluorocarbon, a hydrofluorocarbon, or a hydrocarbon, including trichlorofluoromethane, dichlorodifluoromethane, dichlorotetrafluoroethanol, and 1,1,1,2-tetrafluoroethane, or combinations thereof. Suitable surfactants include sorbitan trioleate and soya lecithin. Oleic acid may also be useful as a surfactant.

Formulations for dispensing from a powder inhaler device will comprise a finely divided dry powder containing protein (or derivative) and may also include a bulking agent, such as lactose, sorbitol, sucrose, mannitol, trehalose, or xylitol in amounts which facilitate dispersal of the powder from the device, e.g., 50 to 90% by weight of the formulation.

Nasal delivery of the protein (or analog or derivative) is also contemplated. Nasal delivery allows the passage of the protein to the blood stream directly after administering the therapeutic product to the nose, without the necessity for deposition of the product in the lung. Formulations for nasal delivery include those with dextran or cyclodextran. Delivery via transport across other mucus membranes is also contemplated.

Dosage

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One skilled in the art will be able to ascertain effective dosages by administration and observing the desired therapeutic effect. Due to the N-terminus modification of the OB protein, the present

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invention provides unexpected protein protection from degradation, and increases circulation time and stability, when compared to OB protein or C-terminus modification of the OB protein. One skilled in the art, therefore, will be able to ascertain from these changes that an effective dosage may require lower doses or less frequent dosing.

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Preferably, the formulation of the molecule will be such that between about .10 μ g/kg/day and 10 mg/kg/day will yield the desired therapeutic effect. 10 The effective dosages may be determined using diagnostic tools over time. For example, a diagnostic for measuring the amount of OB protein or Fc-OB fusion protein in the blood (or plasma or serum) may first be used to determine endogenous levels of protein. Such 15 diagnostic tools may be in the form of an antibody assay, such as an antibody sandwich assay. The amount of endogenous OB protein is quantified initially, and a baseline is determined. The therapeutic dosages are determined as the quantification of endogenous and 20 exogenous OB protein or Fc-OB fusion protein (that is, protein, analog or derivative found within the body, either self-produced or administered) is continued over the course of therapy. The dosages may therefore vary 25 over the course of therapy, with a relatively high dosage being used initially, until therapeutic benefit is seen, and lower dosages used to maintain the therapeutic benefits.

Ideally, in situations where solely reduction
in blood lipid levels is desired, where maintenance of reduction of blood lipid levels is desired, or an increase in lean body mass is desired, the dosage will be insufficient to result in weight loss. Thus, during an initial course of therapy of an obese person, dosages may be administered whereby weight loss and concomitant blood lipid level lowering or concomitant fat tissue

decrease/lean mass increase is achieved. Once sufficient weight loss is achieved, a dosage sufficient to prevent re-gaining weight, yet sufficient to maintain desired blood lipid levels or lean mass increase (or, prevention of lean mass depletion) may be administered. These dosages can be determined empirically, as the effects of OB or Fc-OB protein are reversible, (e.g., Campfield et al., Science 269: 546-549 (1995) at 547). Thus, if a dosage resulting in weight loss is observed when weight loss is not desired, one would administer a lower dose in order to achieve the desired blood lipid levels or increase in lean tissue mass, yet maintain the desired weight.

For increasing an individual's sensitivity to insulin, similar dosage considerations may be taken into account. Lean mass increase without weight loss may be achieved sufficient to decrease the amount of insulin (or, potentially, amylin, thiazolidinediones, or other potential diabetes treating drugs) an individual would be administered for the treatment of diabetes.

For increasing overall strength, there may be similar dosage considerations. Lean mass increase with concomitant increase in overall strength may be achieved with doses insufficient to result in weight loss. Other benefits, such as an increase in red blood cells (and oxygenation in the blood) and a decrease in bone resorption or osteoporosis may also be achieved in the absence of weight loss.

20 <u>Combinations</u>

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The present methods may be used in conjunction with other medicaments, such as those useful for the treatment of diabetes (e.g., insulin, possibly, thiazolidinediones, amylin, or antagonists thereof), cholesterol and blood pressure lowering medicaments (such as those which reduce blood lipid levels or other

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cardiovascular medicaments), and activity increasing medicaments (e.g., amphetamines). Appetite suppressants may also be used (such as those affecting the levels of serotonin or neuropeptide Y). Such administration may be simultaneous or may be <u>in seriatim</u>.

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In addition, the present methods may be used in conjunction with surgical procedures, such as cosmetic surgeries designed to alter the overall appearance of a body (e.g., liposuction or laser surgeries designed to reduce body mass). The health 10 benefits of cardiac surgeries, such as bypass surgeries or other surgeries designed to relieve a deleterious condition caused by blockage of blood vessels by fatty deposits, such as arterial plaque, may be increased with concomitant use of the present compositions and methods. 15 Methods to eliminate gall stones, such as ultrasonic or laser methods, may also be used either prior to, during or after a course of the present therapeutic methods. Furthermore, the present methods may be used as an 20 adjunct to surgeries or therapies for broken bones, damaged muscle, or other therapies which would be improved by an increase in lean tissue mass.

The following examples are offered to more fully illustrate the invention, but are not to be construed as limiting the scope thereof.

EXAMPLE 1: Use of Murine FC-OB Protein Via Subcutaneous Injection

This example demonstrates that injection 30 subcutaneously of murine Fc-OB protein results in weight loss in normal mice. Normal (non-obese) CD1 mice were administered murine Fc-OB protein via subcutaneous injections over a 22 day time period. A dosage of 10 mg protein/kg body weight/day resulted in a 14% (+/- 1.1%) 35 loss from baseline weight by the 22nd day of injections. A dosage of PBS resulted in a 3.9% (+/- 3.3%) loss from

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baseline weight by the 22nd day of injections. weight loss with the use of 10 mg protein/kg body weight/day of Fc-OB protein in obese CD1 mice resulted in a 10% (+/-4.3%) loss from baseline weight and a dosage of PBS resulted in a 8.7% (+/- 1.3%) loss from baseline weight, both by the 22nd day of injections Presented below are the percent (%) differences from baseline weight in CD1 mice (8 weeks old):

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Table 1 Weight Loss Upon Subcutaneous Injection

Time (days)	Vehicle (PBS)	Lean/Recombinant Fc-OB Fusion Protein	Obese/Recombinant Fc-OB Fusion Protein
1-2	44 +/1.1	-3.6 +/41	-1.03 +/- 1.36
3-4	-1.07 +/13	-6.8 +/- 1.5	-2.7 +/- 1.1
5-6	13 +/- 1.1	-9.5 +/- 1.2	-4.9 +/95
7-8	92 +/29	-12.5 +/- 1.6	-7.7 +/- 2.9
9-10	1.6 +/- 1.3	-12.6 +/- 1.9	-8.2 +/- 2.9
11-12	-1.98 +/- 1	-13.6 +/- 1.96	-8.6 +/- 2.9
13-14	-5.2 +/- 1.3	-14.6 +/-1.7	-10.1+/- 3.6
15-16	-8.6 +/- 0.1	-14.5 +/- 2	-9.4 +/- 2.2
17-18	-8.5 +/64	-16.1 +/- 1.8	-9.6 +/- 2.99
19-20	-4.1 +/99	-16 +/- 1.5	-10.4 +/- 3.3
21-22	-3.9 +/- 3.3	-14.1 +/- 1.1	-10 +/- 4.3

15 As can be seen, at the end of a 22 day subcutaneous regime, animals receiving the FC-OB protein lost over 14.1% of their body weight in lean and 10% of body weight in obese, as compared to animals only

Surprisingly, animals receiving Fc-OB protein up to 22 days continued to loose weight up until 28

receiving the PBS vehicle and as compared to baseline.

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days, 4 days after the last injection. Normal (nonobese) CD1 mice administered 10 mg protein/kg body weight/day of murine Fc-OB protein via subcutaneous

injections stopped at day 22 resulted in a 21% loss from baseline weight at day 28 as compared to 14% loss at day 22. Likewise, obese CD1 mice administered 10 mg protein/kg body weight/day of murine Fc-OB protein stopped at day 22 resulted in a 13% loss from baseline weight at day 28 compared to 10% loss at day 22. At day

34 weight loss was maintained at 10% loss in obese mice 10 where lean mice recovered to 5% loss. Controls in each system from day 22 through day 34 averaged from 4% in obese mice and 7% gain in lean mice.

15 EXAMPLE 2: Use of Human FC-OB Protein Via Subcutaneous Injection in C57 Mice

This example demonstrates that injection subcutaneously of human Fc-OB protein results in weight loss in normal mice. Normal (non-obese) C57 mice were 20 administered human Fc-OB protein via subcutaneous injections over a 7 day time period. A dosage of 10 mg protein/kg body weight/day resulted in a 12% (+/- 1.3%) loss from baseline weight by the 7th day of injections. A dosage of 1 mg protein/kg body weight/day resulted in a 8.9% (+/- 1.5%) loss from baseline weight by the 7th 25 day of injections. The weight loss with the use of 10 mg protein/kg body weight/day of human OB protein in obese C57 mice resulted in a 1.1% (+/- .99%) loss from baseline weight and a dosage of 1 mg protein/kg body 30 weight/day resulted in a 2.5% (+/- 1.1%) loss from baseline weight, both by the 7th day of injections.

<u>Results</u>

Presented below are the percent (%)

differences from baseline weight in C57 mice (8 weeks old):

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<u>Table 2</u>					
Weight	Loss	Upon	Subcutaneous	Injection	

Time (days)	Vehicle (PBS)	Recombinant Fc-OB Fusion Protein	Recombinant OB Protein
1-2	.258 +/- 1.3	-6.4 +/- 1.6	-2.1 +/91
3-4	2.2 +/- 1.1	-12.1 +/- 1.5	78 +/36
5-6	4.5 +/- 2	-11.5 +/- 1.5	-1.7 +/6
7-8	7.0 +/- 2.1	-11.9 +/- 1.6	0.1 +/- 1.2
9-10	9.0 +/-1.9	-11.5 +/- 1.3	7.2 +/- 2.7
11-12	10 +/- 3.8	-9 +/-1.4	10.9 +/- 2.9
13-14	12.5 +/- 4.4	-9.5 +/- 1.6	12.3 +/- 6.4
15-16	11.1 +/- 1.0	-3.0 +/- 1.5	10.3 +/- 3.3
17-18	17.2 +/- 3.6	8.0 +/- 1.3	13.3 +/- 3.4

As can be seen, at the end of a day 17 after a 7 day subcutaneous regime at 10 mg/kg/day, animals receiving the FC-OB protein recovered to 8% of their body weight. Animals receiving dosages of 1 mg/kg/day after a 7 day subcutaneous regime returned to 6.4% of body weight after 12 days.

These studies also show that during recovery periods from day 7 to day 22, after the last injection at day 7, body weight recovery is slower in the Fc-OB treated C57 mice that with the OB treated mice. This suggests that the Fc-OB protein is not cleared as quickly as OB protein thereby causing the extended weight loss effect.

EXAMPLE 3: Dose Response of CF7 Mice Treated with Fc-OB Fusion Protein

An additional study demonstrated that there was a dose response to continuous administration of Fc-OB protein. In this study, obese CF7 mice, weighing

35-40 g were administered recombinant human Fc-OB protein using methods similar to the above example. The results are set forth in Table 3, below, (with % body weight lost as compared to baseline, measured as above):

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Table 3

Dose Response With Continuous Administration

Dose	Time	% Reduction in Body Weight
0.25 mg/kg/day	Day 5	4
0.5 mg/kg/day	Day 5	12
1 mg/kg/day	Day 5	16

As can be seen, increasing the dose from 0.25 mg/kg/day to 1 mg/kg/day increased the weight lost from 4% to 16%. It is also noteworthy that at day 5, the 1 mg/kg/day dosage resulted in a 16% reduction in body weight. These studies also showed slow weight recovery rates to 0% suggesting that the Fc-OB protein is not quickly cleared thereby causing the extended weight loss effect.

EXAMPLE 4: Pharmacokinetics of recombinant human Fc-OB in CD-1 Mice and Dogs

This study demonstrated the pharmacokinetic properties of recombinant human met Fc-OB protein in CD-1 mice and dogs. Following intravenous or subcutaneous dosing at 1 mg/kg/day, serum concentrations of recombinant human met Fc-OB protein and human met OB protein were determined by an enzyme-linked immunosorbent assay (ELISA).

In both species, an increase in exposure, as quantified by higher peak serum concentrations and larger areas under-the-serum-concentration-curve (AUC), was observed when compared to recombinant met-human OB

protein. Fc-OB has lower systemic clearance than recombinant met-human OB protein. This is seen in the lower clearance and longer half-life of Fc-OB over OB protein. The increase in size causes not only an increase in protein stability, but also a decrease in the efficiency of renal clearance. As a result, Fc-OB is cleared slower from the systemic circulation. The increases in peak time, peak serum concentrations and AUC for Fc-OB protein are consistent with lower clearance. Fc-OB protein will yield substantially higher systemic exposure when compared to OB protein. Results are shown in Table 4 below:

Table 4
Pharmacokinetic Properties

Species	CD-1	Mice	CD-1	Mice	Beagl	e Dogs
Route of		venous		aneous		aneous
Administration					<u></u>	
	ОВ	Fc-OB	OB	Fc-OB	ОВ	Fc-OB
	protein	protein	protein	protein	protein	protein
Dose Level	1	1	1	1	0.5	0.5
(mg/kg)						
Peak Time (h)			0.14	6	2.8	8
Peak Serum			1520	7550	300	1120
Concentration						
(ng/mL)						
AUC (ng•h/mL)	1470	366000	1230	132000	2200°	52500
Half-life (h)	0.491	21.4	0.388		2.13	22.9
Clearance	681	2.73				
(mL/h/kg)			3.33			

EXAMPLE 5:

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This example demonstrates that in normal mice which are not obese and do not have elevated blood lipid levels, administration of human recombinant Fc-OB

protein results in a lowering of cholesterol, glucose and triglyceride levels. In addition, this example demonstrates that these levels remain low over a three day recovery period.

Normal CD1 mice were administered recombinant human Fc-OB protein via subcutaneous injections. Blood samples were taken 24 hours after day 23, the last day of injection. As discussed above, the animals lost weight at the dosages administered. As shown in Table 5, the mice had substantial reduction of serum cholesterol, glucose and triglycerides in a dosedependent fashion when compared to controls:

Table 5

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Dose	Glucose	Cholesterol	Triglycerides
PBS	232.6 +/- 15.1	67.8 +/- 3.6	52.6 +/- 3.7
1 mg/kg/day	225.8 +/- 29.1	54 +/- 5.6	43 +/- 8.7
10 mg/kg/day	193.2 +/- 21.4	53.4 +/- 5.7	38 +/- 11
1 mg/kg every 2 days	242.0 +/- 9.3	52.6 +/- 4.4	40.8 +/- 7.2
10 mg/kg every 2	197.4 +/- 27.9	51.4 +/- 5.9	29.8 +/- 6.3
days			
1 mg/kg every 3 days	244.8 +/- 19.5	60.8 +/- 7.3	54 +/- 7.1
10 mg/kg every 3	188 +/- 31.2	52.2 +/- 6.9	26.2 +/- 10.7
days			

These data demonstrate that the Fc-OB protein, or analogs or derivatives thereof, are effective blood lipid lowering agents.

EXAMPLE 6:

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A obese human patient is administered human Fc-OB protein, or analog or derivative for the purpose of weight reduction. The obese patient also has elevated

levels of blood lipids, including elevated levels of cholesterol, above 200 mg/100 ml. The patient attains a satisfactory weight reduction over the course of Fc-OB therapy. A maintenance dose of Fc-OB protein or analog or derivative is administered to the non-obese patient to maintain lowered blood lipid levels, including lowered cholesterol levels, below 200 mg/100 ml. The dose administered is insufficient to result in further weight loss. Administration is chronic. Levels of circulating Fc-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody assay against the OB protein (or other antigenic source if applicable).

15 EXAMPLE 7:

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A non-obese human patient undergoes coronary bypass surgery or other invasive treatment to alleviate advanced stages arterial plaque formation. After the surgery, the patient is administered a maintenance dose of Fc-OB protein or analog or derivative in order to prevent the re-formation of arterial plaque. The dose administered is insufficient to result in weight loss. Administration is chronic. Levels of circulating Fc-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody assay against the OB protein (or other antigenic source if applicable).

EXAMPLE 8:

A non-obese human patient experiences

hypertension due to restricted blood flow from clogged arteries. The patient is administered a dose of Fc-OB protein, or analog or derivative thereof sufficient to reduce arterial plaque resulting in clogged arteries.

Thereafter, the patient is monitored for further arterial plaque formation, and hypertension. If the condition re-appears, the patient is re-administered an

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effective amount of Fc-OB protein, analog or derivative sufficient to restore blood flow, yet insufficient to result in weight loss. Levels of circulating Fc-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody assay against the Fc-OB protein (or other antigenic source if applicable).

EXAMPLE 9:

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A human patient experiences gall stones. Either the gall stones are not removed and the formation 10 of additional gall stones is sought to be avoided, or the gall stones are removed but the gall bladder remains (as, for example, using laser or ultrasonic surgery) and the formation of additional gall stones is sought to be 15 avoided. The patient is administered an effective amount of Fc-OB protein, analog or derivative thereof to result in prevention of accumulation of additional gall stones or re-accumulation of gall stones. Levels of circulating Fc-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody 20 assay against the Fc-OB protein (or other antigenic source if applicable).

EXAMPLE 10:

A diabetic human patient desires to use decreased dosages of insulin for treatment of diabetes. The patient is administered an effective amount of Fc-OB protein, analog or derivative thereof to result in an increase in lean tissue mass. The patient's sensitivity to insulin increases, and the dosage of insulin necessary to alleviate symptoms of diabetes is decreased, either in terms of a decrease in the units of insulin needed, or in terms of a decrease in the number of injections of insulin needed per day. Levels of circulating Fc-OB protein or analog or derivative may be

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monitored using a diagnostic kit, such as an antibody assay against the OB protein (or other antigenic source if applicable).

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5 EXAMPLE 11:

A non-obese human patient desires an increase in lean tissue mass for therapeutic purposes, such as recovery from illness which depleted lean tissue mass. The patient is administered an effective amount of Fc-OB 10 protein, analog or derivative thereof to result in the desired increase in lean tissue mass. Increase in lean tissue mass is monitored using DEXA scanning. Levels of circulating Fc-OB protein or analog or derivative may be monitored using a diagnostic kit, such as an antibody assay against the OB protein (or other antigenic source 15 if applicable).

MATERIALS AND METHODS

- 20 Animals. Wild type CD1 mice and (+/+) C57B16 mice were used for the above examples. The age of the mice at the initial time point was 8 weeks, and the animals were weight stabilized.
- 25 Feeding and Weight Measurement. Mice were given ground rodent chow (PMI Feeds, Inc.) in powdered food feeders (Allentown Caging and Equipment) which allowed a more accurate and sensitive measurement than use of regular block chow. Weight was measured at the 30 same time each day (2:00 p.m.), for the desired period. Body weight on the day prior to the injection was defined as baseline weight. The mice used weighed 18-22 grams.
- 35 Housing. Mice were single-housed, and maintained under humane conditions.

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Administration of Protein or Vehicle. Protein (as described below) or vehicle (phosphate buffered saline, pH 7.4) were administered by subcutaneous injections or intravenously.

Controls. Control animals were those who were injected with the vehicle alone without either Fc-OB fusion protein or OB protein added to the vehicle.

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Protein. Sequence ID. Nos. 1, 2 and 3 set forth murine recombinant OB DNA and protein (Figure 1), and Sequence ID. Nos. 4, 5 and 6 set forth an analog recombinant human OB DNA and protein (Figure 2). As noted above recombinant human OB protein as in SEQ. ID.. NO. 6 has a lysine residue at position 35 and an isoleucine residue at position 74. Furthermore, the recombinant human protein set forth in Zhang et al., Nature, supra, and PCT publication WO 96/05309 (12/22/96) (both incorporated by reference including figures), and the murine and human analog recombinant proteins of Figures 1 and 2 are illustrative of the OB protein which may be used in forming the Fc-OB fusion protein of the present methods of treatment and manufacture of a medicament. Other OB or Fc proteins or analogs or derivatives thereof may also be used to form the Fc-OB fusion protein.

Herein, the first amino acid of the amino acid sequence for recombinant OB protein is referred to as +1, and is valine, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 146 (cysteine) (see Figures 1 and 2). The first amino acid sequence for recombinant human Fc-OB protein of Figure 3 is referred to as +1, and is glutamate, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 378 (cysteine). The first amino acid sequence for the recombinant human Fc-OB protein variant

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of Figure 4 is referred to as +1, and is glutamate, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 378 (cysteine). The first amino acid sequence for the recombinant human Fc-OB protein variant of Figure 5 is referred to as +1, and is aspartic acid, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 373 (cysteine). The first amino acid sequence for the recombinant human Fc-OB protein variant of Figure 6 is referred to as +1, and is aspartic acid, and the amino acid at position -1 is methionine. The C-terminal amino acid is number is 373 (cysteine).

Expression Vector and Host Strain

15 The plasmid expression vector used is pAMG21
(ATCC accession number 98113), which is a derivative of
pCFM1656 (ATCC accession number 69576) and contains
appropriate restriction sites for insertion of genes
downstream from the lux PR promoter (see US Patent No.
20 5,169,318 for a description of the lux expression
system). The Fc-OB DNA, described below and shown in
Figures 3-6, was created and ligated into the expression

NdeI and BamHI and transformed into the <u>E. coli</u> host strain, FM5. <u>E. coli</u> FM5 cells were derived at Amgen Inc., Thousand Oaks, CA from <u>E. coli</u> K-12 strain (Bachmann, et al., Bacterial. Rev. <u>40</u>: 116-167 (1976)) and contain the integrated lambda phage repressor gene, cI857 (Sussman et al., C. R. Acad. Sci. <u>254</u>: 1517-1579

vector pAMG21 linearized with restriction endonucleases

(1962)). Vector production, cell transformation, and colony selection were performed by standard methods, (e.g., Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2d Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.) Host cells were grown in LB media.

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Fc-OB DNA Construction

The plasmid pFc-A3 (described below) served as the source of sequence for human immunoglobulin IgG-1 heavy chain from amino acid number 99 (Glu) to the natural carboxyl terminus. The human IgG-1 sequence can be obtained from Genebank (P01857).

The human OB sequence is disclosed above as well as Zhang et al., Nature, supra, and PCT publication WO 96/05309 both incorporated by reference including drawings. The OB DNA was ligated into the expression vector pCFM1656 linearized with restriction endonucleases XbaI and BamHI using standard cloning procedures, e.g., Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2d Edition, Cold Spring Harbor
Laboratory Press, Cold Spting Harbor, N.Y.. The plasmid pCFM1656 carrying the OB DNA sequence served as the source of sequence for the recombinant human OB gene.

The genetic fusing of these two sequences was carried out by the method of PCR overlap extension (Ho, S.N., et al., Site Directed Mutagenesis By Overlap 20 Extension Using The Polymerase Chain Reaction, Gene 77:51-59(1989)). The product of the PCR was cleaved with restriction endonuclease NdeI to create a 5'-cohesive end and with restriction endonuclease BamHI to create a 3'-cohesive terminus. The vector, pAMG21, was similarly cleaved. A ligation was performed with the fusion fragment and the linearized vector. Ligated DNA was transformed by electroporation into the E. coli host strain. Clones surviving on kanamycin (50µg/ml) 30 selection agar plates were checked for expression of Fc-OB-sized protein. Plasmid from individual clones was isolated and the sequence of the gene coding region verified.

When additional modifications of the Fc-OB gene were desired, the PCR technique was used again to engineer the changes. Two sets of changes were -44-

performed at the N-terminus of the Fc portion of the fusion protein (SEQ. ID. No. 9) to create the variants SEQ. ID. NOS. 12 and 15. Another variant was constructed to introduce four amino acid substitutions to ablate the Fc-receptor binding site (leucine at position 15 substituted with glutamate), and the complement (Clq) binding site (glutamate at position 98 substituted with alanine, lysine at position 100 substituted with alanine, and lysine at position 102 substituted with alanine (See, Xin Xiao Zheng et. al, J. Immunol. 154:5590-5600 (1995)). The template for this construct was Seq. ID. No. 15 and the resulting variant was SEQ. ID. Nos. 18.

15 pFC-A3 Vector Construction

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A plasmid, pFc-A3, containing the region encoding the Fc portion of human immunoglobulin IgG-1 heavy chain (See Ellison, J. W. et. al, Nucleic Acids Res. 10:4071-4079 (1982)), from the first amino acid 20 Glu-99 of the hinge domain to the carboxyl terminus plus a 5'-NotI fusion site and 3'-SalI and XbaI sites, was made by PCR amplification of the human spleen cDNA library. PCR reactions were in a final volume of 100 ml and employed 2 units of Vent DNA polymerase in 20 mM 25 Tris-HCl (pH 8.8), 10 mM KCl, 10 mM (NH4)2SO4, 2 mM MgSO4, 0.1% Triton X-100 with 400 mM each dNTP and 1 ng of the cDNA library to be amplified together with 1 uM of each primer. Reactions were initiated by denaturation at 95 °C for 2 min, followed by 30 cycles of 95 °C for 30 s, 55 $^{\circ}$ C for 30 s, and 73 $^{\circ}$ C for 2 min. The 5'-30 primer incorporated a NotI site immediately 5' to the first residue (Glu-99) of the hinge domain of IgG-1. The 3'-primer incorporated SalI and XbaI sites. base pair PCR product was digested with NotI and SalI, the resulting DNA fragment was isolated by

electrophoresis through 1% agarose and purified and

cloned into NotI, SalI-digested pBluescript II KS vector (Stratagene). The insert in the resulting plasmid, pFc-A3, was sequenced to confirm the fidelity of the PCR reaction.

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Methods for Production

The methods below for production have been used to produce biologically active recombinant methionyl murine or human analog OB protein and Fc-OB fusion proteins. Similar methods may be used to prepare biologically active methionyl human OB protein.

Fermentation Process

A batch fermentation process was used. Media compositions are set forth below.

A portion of the media consisting of primarily nitrogen sources was sterilized (by raising temperature to 120~123°C for 25~35 minutes) in the fermentation vessel. Upon cooling, carbon, magnesium, phosphate, and trace metal sources were added aseptically. An overnight culture of the above recombinant murine protein-producing bacteria of 500 mL (grown in LB broth) was added to the fermentor. When the culture optical density (measured at 600 nm as an indicator for cell density) reached 15~25 absorption units, an autoinducer solution (0.5 mg/mL homoserine lactone) was added (1 mL/L) to the culture to induce the recombinant gene expression. The fermentation process was allowed to continue for additional 10 to 16 hours, followed by harvesting the broth by centrifugation.

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Media Composition:

Batch: 34 g/L Yeast extract 78 g/L Soy peptone 0.9 g/LPotassium chloride 5 5.0 g/LHexaphos 1.7 g/L Citric acid 120 g/L Glycerol 0.5 g/L MgSO $4 \cdot 7H_2O$ 0.2 mL/L Trace Metal Solution 10 0.5 mL/L P2000 Antifoam

Trace Metal Solution:

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Ferric Chloride (FeCl3·6H2O): 27 g/L Zinc Chloride (ZnCl₂·4H₂O): 2 g/L 15 Cobalt Chloride (CoCl₂·6H₂O): 2 g/L Sodium Molybdate (NaMoO4·2H2O): 2 g/L Calcium Chloride (CaCl₂·2H₂O): 1 g/L Cupric Sulfate (CuSO4.5H2O): 1.9 g/L Boric Acid (H3BO3): 0.5 g/L 20 Manganese Chloride (MnCl2·4H2O): 1.6 g/L Sodium Citrate dihydrate: 73.5 g/L

Purification Process for Human Fc-OB Fusion Protein

Purification for human Fc-OB fusion protein was accomplished by the steps below (unless otherwise noted, the following steps were performed at 4'C). Purification for murine and human OB protein is disclosed in PCT publication WO 96/05309, supra, herein incorporated by reference.

1. Cell paste. E. coli cell paste was suspended in 5 times volumes of distilled water. The cells in the water were further broken by two passes

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through a microfluidizer. The broken cells were centrifuged at 4.2k rpm for 1 hour in a Beckman JB-6 centrifuge with a J5-4.2 rotor.

- 2. Inclusion body wash. The supernatant from above was removed and the pellet was resuspended with five volumes of distilled water. The mixture was centrifuged as in step 1.
- 3. Solubilization. The pellet was solubilized with 10 volumes of 50 mM tris, pH 8.5, 8 M guanidine hydrochloride, 10 mM dithiothreitol and stirred for one 10 hour at room temperature. The solution is made 40 mM cystamine dihydrochloride and stirred for one hour.
 - 4. The solution from step 3 is added to 20 to 30 volumes of the following refold solution: 50 mM tris, pH 8.5, 0.8 M arginine, 2 M urea, and 4 mM cysteine. The refold is stirred for 16 hours at 8°C.
 - 5. Buffer exchange. The solution from step 4 is concentrated and diafiltered into 10 mM tris, pH 8.5.
- 6. Acid precipitation. The solution from step 5 is adjusted to pH 4.75 with 50% glacial acid and 20 incubated for 30 minutes at room temperature. The solution is filtered.
 - 7. Cation exchange chromatography. The solution from step 6 is adjusted to pH 7.0 and loaded onto a CM Sepharose Fast Flow column at 10'C. A twenty column volume gradient is done at 10 mM phosphate, pH 7.0, 0 to 0.1 M NaCl.

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- 8. Anion exchange chromatography. The CM elution pool from step 7 is diluted 5 fold with 5 mM 30 tris, pH 7.5 and loaded onto a Q Sepharose Fast Flow at 10'C. A 20 column volume gradient is done at 10 mM tris, pH 7.5, 0 to 0.2M NaCl.
- 9. Hydrophobic interaction chromatography. The Q sepharose pool is made 0.75M ammonium sulfate and 35 loaded on a methyl Macroprep hydrophobic interaction

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column at room temperature. A 20 column volume gradient is done at 10 mM phosphate, pH 7.0, 0.75M to 0M ammonium sulfate.

10. Buffer exchange. The pool from step 9 is concentrated as necessary and dialyzed against PBS buffer.

While the present invention has been described in terms of preferred embodiments, it is understood that variations and modifications will occur to those skilled in the art. Therefore, it is intended that the appended claims cover all such equivalent variations which come within the scope of the invention as claimed.

15

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Mann, Michael B. Hecht, Randy I.
- (ii) TITLE OF INVENTION: OB FUSION PROTEIN COMPOSITIONS AND METHODS
- (iii) NUMBER OF SEQUENCES: 18
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Amgen Inc.
 - (B) STREET: 1840 DeHavilland Drive
 - (C) CITY: Thousand Oaks
 - (D) STATE: CA
 - (E) COUNTRY: USA
 - (F) ZIP: 91320-1789
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/770,973
 - (B) FILING DATE: 20-DEC-1996
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Knight, Matthew W.
 - (B) REGISTRATION NUMBER: 36,846
 - (C) REFERENCE/DOCKET NUMBER: A-416
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 491 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION: 41
 - (D) OTHER INFORMATION: /note= "Met = ATG"

- 50 -

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, ,			360 :0	147.7

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TCAGGACGAC	ACCAAAACCT	TAATTAAAAC	GATCGTTACG	CGTATCAACG	ACATCAGTCA	120
CACCCAGTCG	GTCTCCGCTA	AACAGCGTGT	TACCGGTCTG	GACTTCATCC	CGGGTCTGCA	180
CCCGATCCTA	AGCTTGTCCA	AAATGGACCA	GACCCTGGCT	GTATACCAGC	AGGTGTTAAC	240
CTCCCTGCCG	TCCCAGAACG	TTCTTCAGAT	CGCTAACGAC	CTCGAGAACC	TTCGCGACCT	300
GCTGCACCTG	CTGGCATTCT	CCAAATCCTG	CTCCCTGCCG	CAGACCTCAG	GTCTTCAGAA	360
ACCGGAATCC	CTGGACGGG	TCCTGGAAGC	ATCCCTGTAC	AGCACCGAAG	TTGTTGCTCT	420
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(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 491 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

AGATCTAAAC	TCAAAATTGA	AAATCTTCCT	CCTTATTGTA	TACCATGGCT	AGGTCTTTCA	60
AGTCCTGCTG	TGGTTTTGGA	ATTAATTTTG	CTAGCAATGC	GCATAGTTGC	TGTAGTCAGT	120
GTGGGTCAGC	CAGAGGCGAT	TTGTCGCACA	ATGGCCAGAC	CTGAAGTAGG	GCCCAGACGT	180
GGGCTAGGAT	TCGAACAGGT	TTTACCTGGT	CTGGGACCGA	CATATGGTCG	TCCACAATTG	240
GAGGGACGGC	AGGGTCTTGC	AAGAAGTCTA	GCGATTGCTG	GAGCTCTTGG	AAGCGCTGGA	300
CGACGTGGAC	GACCGTAAGA	GGTTTAGGAC	GAGGGACGGC	GTCTGGAGTC	CAGAAGTCTT	360
TGGCCTTAGG	GACCTGCCCC	AGGACCTTCG	TAGGGACATG	TCGTGGCTTC	AACAACGAGA	420
CAGGCCAGAC	GTCCCAAGGG	AAGTCCTGTA	GGAAGTCGTC	GACCTGCAAA	GAGGCCTTAC	480
AATTACCTAG	G	•				491

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(2) INFORMATION FOR SEQ ID NO: 3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 147 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: protein
- (ix) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "Met (ATG) starts at -1"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Val Pro Ile Gln Lys Val Gln Asp Asp Thr Lys Thr Leu Ile Lys 15

Thr Ile Val Thr Arg Ile Asn Asp Ile Ser His Thr Gln Ser Val Ser 20 25

Ala Lys Gln Arg Val Thr Gly Leu Asp Phe Ile Pro Gly Leu His Pro 40

Ile Leu Ser Leu Ser Lys Met Asp Gln Thr Leu Ala Val Tyr Gln Gln 55

Val Leu Thr Ser Leu Pro Ser Gln Asn Val Leu Gln Ile Ala Asn Asp 70 75

Leu Glu Asn Leu Arg Asp Leu Leu His Leu Leu Ala Phe Ser Lys Ser

Cys Ser Leu Pro Gln Thr Ser Gly Leu Gln Lys Pro Glu Ser Leu Asp 105

Gly Val Leu Glu Ala Ser Leu Tyr Ser Thr Glu Val Val Ala Leu Ser 115 120 125

Arg Leu Gln Gly Ser Leu Gln Asp Ile Leu Gln Gln Leu Asp Val Ser 130 135

Pro Glu Cys

145

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(2)	INFORMATION	FOR SEQ	ID	NO: 4	:
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	(i)	SEQUENCE	CHARACTERISTICS	:
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(A) LENGTH: 454 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 4
- (D) OTHER INFORMATION: /note= "Met = ATG"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CATATGGTAC	CGATCCAGAA	AGTTCAGGAC	GACACCAAAA	CCTTAATTAA	AACGATCGTT	60
ACGCGTATCA	ACGACATCAG	TCACACCCAG	TCGGTGAGCT	CTAAACAGCG	TGTTACAGGC	120
CTGGACTTCA	TCCCGGGTCT	GCACCCGATC	CTGACCTTGT	CCAAAATGGA	CCAGACCCTG	180
GCTGTATACC	AGCAGATCTT	AACCTCCATG	CCGTCCCGTA	ACGTTCTTCA	GATCTCTAAC	240
GACCTCGAGA	ACCTTCGCGA	CCTGCTGCAC	GTGCTGGCAT	TCTCCAAATC	CTGCCACCTG	300
CCATGGGCTT	CAGGTCTTGA	GACTCTGGAC	TCTCTGGGCG	GGGTCCTGGA	AGCATCCGGT	360
TACAGCACCG	AAGTTGTTGC	TCTGTCCCGT	CTGCAGGGTT	CCCTTCAGGA	CATGCTTTGG	420
CAGCTGGACC	TGTCTCCGGG	TTGTTAATGG	ATCC			454

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 454 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

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GACCTGAAGT	AGGGCCCAGA	CGTGGGCTAG	GACTGGAACA	GGTTTTACCT	GGTCTGGGAC	180
CGACATATGG	TCGTCTAGAA	TTGGAGGTAC	GGCAGGGCAT	TGCAAGAAGT	CTAGAGATTG	240
CTGGAGCTCT	TGGAAGCGCT	GGACGACGTG	CACGACCGTA	AGAGGTTTAG	GACGGTGGAC	300
GGTACCCGAA	GTCCAGAACT	CTGAGACCTG	AGAGACCCGC	CCCAGGACCT	TCGTAGGCCA	360
ATGTCGTGGC	TTCAACAACG	AGACAGGGCA	GACGTCCCAA	GGGAAGTCCT	GTACGAAACC	420
GTCGACCTGG	ACAGAGGCCC	AACAATTACC	TAGG			454

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 147 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: protein
- (ix) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "Met (ATG) starts at -1"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Val Pro Ile Gln Lys Val Gln Asp Asp Thr Lys Thr Leu Ile Lys

1 10 15

Thr Ile Val Thr Arg Ile Asn Asp Ile Ser His Thr Gln Ser Val Ser 20 25 30

Ser Lys Gln Arg Val Thr Gly Leu Asp Phe Ile Pro Gly Leu His Pro 35 40 45

Ile Leu Thr Leu Ser Lys Met Asp Gln Thr Leu Ala Val Tyr Gln Gln 50 55 60

Ile Leu Thr Ser Met Pro Ser Arg Asn Val Leu Gln Ile Ser Asn Asp 65 70 75 80

Leu Glu Asn Leu Arg Asp Leu Leu His Val Leu Ala Phe Ser Lys Ser 85 90 95

Cys His Leu Pro Trp Ala Ser Gly Leu Glu Thr Leu Asp Ser Leu Gly 100 105 110

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Gly Val Leu Glu Ala Ser Gly Tyr Ser Thr Glu Val Val Ala Leu Ser 120

Arg Leu Gln Gly Ser Leu Gln Asp Met Leu Trp Gln Leu Asp Leu Ser 135

Pro Gly Cys 145

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1150 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION: 4
 - (D) OTHER INFORMATION: /note= "Met = ATG"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CATATGGAAC CCAAATCTTG TGACAAAACT CACACATGCC CACCGTGCCC AGCACCTGAA 60 CTCCTGGGGG GACCGTCAGT CTTCCTCTTC CCCCCAAAAC CCAAGGACAC CCTCATGATC 120 TCCCGGACCC CTGAGGTCAC ATGCGTGGTG GTGGACGTGA GCCACGAAGA CCCTGAGGTC 180 AAGTTCAACT GGTACGTGGA CGGCGTGGAG GTGCATAATG CCAAGACAAA GCCGCGGGAG 240 GAGCAGTACA ACAGCACGTA CCGTGTGGTC AGCGTCCTCA CCGTCCTGCA CCAGGACTGG 300 CTGAATGGCA AGGAGTACAA GTGCAAGGTC TCCAACAAAG CCCTCCCAGC CCCCATCGAG 360 AAAACCATCT CCAAAGCCAA AGGGCAGCCC CGAGAACCAC AGGTGTACAC CCTGCCCCCA 420 TCCCGGGATG AGCTGACCAA GAACCAGGTC AGCCTGACCT GCCTGGTCAA AGGCTTCTAT 480 CCCAGCGACA TCGCCGTGGA GTGGGAGAGC AATGGGCAGC CGGAGAACAA CTACAAGACC 540 ACGCCTCCCG TGCTGGACTC CGACGGCTCC TTCTTCCTCT ACAGCAAGCT CACCGTGGAC 600 AAGAGCAGGT GGCAGCAGGG GAACGTCTTC TCATGCTCCG TGATGCATGA GGCTCTGCAC 660 AACCACTACA CGCAGAAGAG CCTCTCCCTG TCTCCGGGTA AAGTACCGAT CCAGAAAGTT 720 ·

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CAGGACG	ACA	CCAAAACCTT	AATTAAAACG	ATCGTTACGC	GTATCAACGA	CATCAGTCAC	780
ACCCAGI	CGG	TGAGCTCTAA	ACAGAAAGTT	ACAGGCCTGG	ACTTCATCCC	GGGTCTGCAC	840
CCGATCC	TGA	CCTTGTCCAA	AATGGACCAG	ACCCTGGCTG	TATACCAGCA	GATCTTAACC	900
TCCATGO	CGT	CCCGTAACGT	TATCCAGATC	TCTAACGACC	TCGAGAACCT	TCGCGACCTG	960
CTGCACG	TGC	TGGCATTCTC	CAAATCCTGC	CACCTGCCAT	GGGCTTCAGG	TCTTGAGACT	1020
CTGGACT	CTC	TGGGCGGGGT	CCTGGAAGCA	TCCGGTTACA	GCACCGAAGT	TGTTGCTCTG	1080
TCCCGTC	TGC	AGGGTTCCCT	TCAGGACATG	CTTTGGCAGC	TGGACCTGTC	TCCGGGTTGT	1140
TAATGGA	TCC						1150

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1150 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GTATACCTTG GGTTTAGAAC ACTGTTTTGA GTGTGTACGG GTGGCACGGG TCGTGGACTT 60 GAGGACCCCC CTGGCAGTCA GAAGGAGAAG GGGGGTTTTG GGTTCCTGTG GGAGTACTAG 120 AGGGCCTGGG GACTCCAGTG TACGCACCAC CACCTGCACT CGGTGCTTCT GGGACTCCAG 180 TTCAAGTTGA CCATGCACCT GCCGCACCTC CACGTATTAC GGTTCTGTTT CGGCGCCCTC 240 CTCGTCATGT TGTCGTGCAT GGCACACCAG TCGCAGGAGT GGCAGGACGT GGTCCTGACC 300 GACTTACCGT TCCTCATGTT CACGTTCCAG AGGTTGTTTC GGGAGGGTCG GGGGTAGCTC 360 TTTTGGTAGA GGTTTCGGTT TCCCGTCGGG GCTCTTGGTG TCCACATGTG GGACGGGGGT 420 AGGGCCCTAC TCGACTGGTT CTTGGTCCAG TCGGACTGGA CGGACCAGTT TCCGAAGATA 480 GGGTCGCTGT AGCGGCACCT CACCCTCTCG TTACCCGTCG GCCTCTTGTT GATGTTCTGG 540 TGCGGAGGC ACGACCTGAG GCTGCCGAGG AAGAAGGAGA TGTCGTTCGA GTGGCACCTG 600 TTCTCGTCCA CCGTCGTCCC CTTGCAGAAG AGTACGAGGC ACTACGTACT CCGAGACGTG 660 TTGGTGATGT GCGTCTTCTC GGAGAGGGAC AGAGGCCCAT TTCATGGCTA GGTCTTTCAA 720

GTCCTGCTGT	GGTTTTGGAA	TTAATTTTGC	TAGCAATGCG	CATAGTTGCT	GTAGTCAGTG	780
TGGGTCAGCC	ACTCGAGATT	TGTCTTTCAA	TGTCCGGACC	TGAAGTAGGG	CCCAGACGTG	840
GGCTAGGACT	GGAACAGGTT	TTACCTGGTC	TGGGACCGAC	ATATGGTCGT	CTAGAATTGG	900
AGGTACGGCA	GGGCATTGCA	ATAGGTCTAG	AGATTGCTGG	AGCTCTTGGA	AGCGCTGGAC	960
GACGTGCACG	ACCGTAAGAG	GTTTAGGACG	GTGGACGGTA	CCCGAAGTCC	AGAACTCTGA	1020
GACCTGAGAG	ACCCGCCCCA	GGACCTTCGT	AGGCCAATGT	CGTGGCTTCA	ACAACGAGAC	1080
AGGGCAGACG	TCCCAAGGGA	AGTCCTGTAC	GAAACCGTCG	ACCTGGACAG	AGGCCCAACA	1140
ATTACCTAGG						1150

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 379 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: protein
 - (ix) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "Met (ATG) starts at -1"
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro 1 5 10 15

Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys 20 25 30

Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val 35 40 45

Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr 50 55 60

Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu 65 70 75 80

- Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His 85 90 95
- Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys
 100 105 110
- Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln
 115 120 125
- Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu 130 135 140
- Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro 145 150 155 160
- Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn 165 170 175
- Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu 180 185 190
- Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val 195 200 205
- Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln 210 215 220
- Lys Ser Leu Ser Leu Ser Pro Gly Lys Val Pro Ile Gln Lys Val Gln 225 230 240
- Asp Asp Thr Lys Thr Leu Ile Lys Thr Ile Val Thr Arg Ile Asn Asp 245 250 255
- Ile Ser His Thr Gln Ser Val Ser Ser Lys Gln Lys Val Thr Gly Leu 260 265 270
- Asp Phe Ile Pro Gly Leu His Pro Ile Leu Thr Leu Ser Lys Met Asp 275 280 285
- Gln Thr Leu Ala Val Tyr Gln Gln Ile Leu Thr Ser Met Pro Ser Arg 290 295 300
- Asn Val Ile Gin Ile Ser Asn Asp Leu Glu Asn Leu Arg Asp Leu Leu 305 310 315 320
- His Val Leu Ala Phe Ser Lys Ser Cys His Leu Pro Trp Ala Ser Gly 325 330 335
- Leu Glu Thr Leu Asp Ser Leu Gly Gly Val Leu Glu Ala Ser Gly Tyr 340 345 350
- Ser Thr Glu Val Val Ala Leu Ser Arg Leu Gln Gly Ser Leu Gln Asp 355 360 365

Met Leu Trp Gln Leu Asp Leu Ser Pro Gly Cys 370 375

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1150 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 4
- (D) OTHER INFORMATION: /note= "Met = ATG"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CATATGGAAC CAAAATCTGC TGACAAAACT CACACATGTC CACCTTGTCC AGCTCCGGAA 60 CTCCTGGGGG GTCCTTCAGT CTTCCTCTTC CCCCCAAAAC CCAAGGACAC CCTCATGATC 120 TCCCGGACCC CTGAGGTCAC ATGCGTGGTG GTGGACGTGA GCCACGAAGA CCCTGAGGTC 180 AAGTTCAACT GGTACGTGGA CGGCGTGGAG GTGCATAATG CCAAGACAAA GCCGCGGGAG 240 GAGCAGTACA ACAGCACGTA CCGTGTGGTC AGCGTCCTCA CCGTCCTGCA CCAGGACTGG 300 CTGAATGGCA AGGAGTACAA GTGCAAGGTC TCCAACAAAG CCCTCCCAGC CCCCATCGAG 360 AAAACCATCT CCAAAGCCAA AGGGCAGCCC CGAGAACCAC AGGTGTACAC CCTGCCCCCA 420 TCCCGGGATG AGCTGACCAA GAACCAGGTC AGCCTGACCT GCCTGGTCAA AGGCTTCTAT 480 CCCAGCGACA TCGCCGTGGA GTGGGAGAGC AATGGGCAGC CGGAGAACAA CTACAAGACC 540 ACGCCTCCCG TGCTGGACTC CGACGGCTCC TTCTTCCTCT ACAGCAAGCT CACCGTGGAC 600 AAGAGCAGGT GGCAGCAGGG GAACGTCTTC TCATGCTCCG TGATGCATGA GGCTCTGCAC 660 AACCACTACA CGCAGAAGAG CCTCTCCCTG TCTCCGGGTA AAGTACCGAT CCAGAAAGTT 720 CAGGACGACA CCAAAACCTT AATTAAAACG ATCGTTACGC GTATCAACGA CATCAGTCAC 780 ACCCAGTCGG TGAGCTCTAA ACAGAAAGTT ACAGGCCTGG ACTTCATCCC GGGTCTGCAC 900 CCCATCCTCA COTTCTCCAA AATCGACCAG ACCCTGGCTG TATACCAGCA GATCTTAACC

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TCCATGCCGT	CCCGTAACGT	TATCCAGATC	TCTAACGACC	TCGAGAACCT	TCGCGACCTG	960
CTGCACGTGC	TGGCATTCTC	CAAATCCTGC	CACCTGCCAT	GGGCTTCAGG	TCTTGAGACT	1020
CTGGACTCTC	TGGGCGGGGT	CCTGGAAGCA	TCCGGTTACA	GCACCGAAGT	TGTTGCTCTG	1080
TCCCGTCTGC	AGGGTTCCCT	TCAGGACATG	CTTTGGCAGC	TGGACCTGTC	TCCGGGTTGT	1140
TAATGGATCC						1150

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1150 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GTATACCTTG GTTTTAGACG ACTGTTTTGA GTGTGTACAG GTGGAACAGG TCGAGGCCTT 60 GAGGACCCC CAGGAAGTCA GAAGGAGAAG GGGGGTTTTG GGTTCCTGTG GGAGTACTAG 120 AGGGCCTGGG GACTCCAGTG TACGCACCAC CACCTGCACT CGGTGCTTCT GGGACTCCAG 180 TTCAAGTTGA CCATGCACCT GCCGCACCTC CACGTATTAC GGTTCTGTTT CGGCGCCCTC 240 CTCGTCATGT TGTCGTGCAT GGCACACCAG TCGCAGGAGT GGCAGGACGT GGTCCTGACC 300 GACTTACCGT TCCTCATGTT CACGTTCCAG AGGTTGTTTC GGGAGGGTCG GGGGTAGCTC 360 TTTTGGTAGA GGTTTCGGTT TCCCGTCGGG GCTCTTGGTG TCCACATGTG GGACGGGGT 420 AGGGCCCTAC TCGACTGGTT CTTGGTCCAG TCGGACTGGA CGGACCAGTT TCCGAAGATA 480 GGGTCGCTGT AGCGGCACCT CACCCTCTCG TTACCCGTCG GCCTCTTGTT GATGTTCTGG 540 TGCGGAGGC ACGACCTGAG GCTGCCGAGG AAGAAGGAGA TGTCGTTCGA GTGGCACCTG 600 TTCTCGTCCA CCGTCGTCCC CTTGCAGAAG AGTACGAGGC ACTACGTACT CCGAGACGTG 660 TTGGTGATGT GCGTCTTCTC GGAGAGGGAC AGAGGCCCAT TTCATGGCTA GGTCTTTCAA 720 GTCCTGCTGT GGTTTTGGAA TTAATTTTGC TAGCAATGCG CATAGTTGCT GTAGTCAGTG 780 TGGGTCAGCC ACTCGAGATT TGTCTTTCAA TGTCCGGACC TGAAGTAGGG CCCAGACGTG

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GGCTAGGACT	GGAACAGGTT	TTACCTGGTC	TGGGACCGAC	ATATGGTCGT	CTAGAATTGG	900
AGGTACGGCA	GGGCATTGCA	ATAGGTCTAG	AGATTGCTGG	AGCTCTTGGA	AGCGCTGGAC	960
GACGTGCACG	ACCGTAAGAG	GTTTAGGACG	GTGGACGGTA	CCCGAAGTCC	AGAACTCTGA	1020
GACCTGAGAG	ACCCGCCCCA	GGACCTTCGT	AGGCCAATGT	CGTGGCTTCA	ACAACGAGAC	1080
AGGGCAGACG	TCCCAAGGGA	AGTCCTGTAC	GAAACCGTCG	ACCTGGACAG	AGGCCCAACA	1140
ATTACCTAGG						1150

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 379 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: protein
- (ix) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "Met (ATG) starts at -1"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Glu Pro Lys Ser Ala Asp Lys Thr His Thr Cys Pro Pro Cys Pro 1 5 10 15

Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys
20 25 30

Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val 35 40 45

Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr 50 55 60

Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu 65 70 75 80

Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His
85 90 95

Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys
100 105 110

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Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln

Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu

Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro

Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn

Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu

Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val

Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln

Lys Ser Leu Ser Pro Gly Lys Val Pro Ile Gln Lys Val Gln

Asp Asp Thr Lys Thr Leu Ile Lys Thr Ile Val Thr Arg Ile Asn Asp

Ile Ser His Thr Gln Ser Val Ser Ser Lys Gln Lys Val Thr Gly Leu

Asp Phe Ile Pro Gly Leu His Pro Ile Leu Thr Leu Ser Lys Met Asp

Gln Thr Leu Ala Val Tyr Gln Gln Ile Leu Thr Ser Met Pro Ser Arg

Asn Val Ile Gln Ile Ser Asn Asp Leu Glu Asn Leu Arg Asp Leu Leu

His Val Leu Ala Phe Ser Lys Ser Cys His Leu Pro Trp Ala Ser Gly

Leu Glu Thr Leu Asp Ser Leu Gly Gly Val Leu Glu Ala Ser Gly Tyr

Ser Thr Glu Val Val Ala Leu Ser Arg Leu Gln Gly Ser Leu Gln Asp

Met Leu Trp Gln Leu Asp Leu Ser Pro Gly Cys

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(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1135 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 4
- (D) OTHER INFORMATION: /note= "Met = ATG"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CATATGGACA AAACTCACAC ATGTCCACCT TGTCCAGCTC CGGAACTCCT GGGGGGTCCT 60 TCAGTCTTCC TCTTCCCCCC AAAACCCAAG GACACCCTCA TGATCTCCCG GACCCCTGAG 120 GTCACATGCG TGGTGGTGGA CGTGAGCCAC GAAGACCCTG AGGTCAAGTT CAACTGGTAC 180 GTGGACGCG TGGAGGTGCA TAATGCCAAG ACAAAGCCGC GGGAGGAGCA GTACAACAGC 240 ACGTACCGTG TGGTCAGCGT CCTCACCGTC CTGCACCAGG ACTGGCTGAA TGGCAAGGAG 300 TACAAGTGCA AGGTCTCCAA CAAAGCCCTC CCAGCCCCCA TCGAGAAAAC CATCTCCAAA 360 GCCAAAGGGC AGCCCCGAGA ACCACAGGTG TACACCCTGC CCCCATCCCG GGATGAGCTG 420 ACCAAGAACC AGGTCAGCCT GACCTGCCTG GTCAAAGGCT TCTATCCCAG CGACATCGCC 480 GTGGAGTGGG AGAGCAATGG GCAGCCGGAG AACAACTACA AGACCACGCC TCCCGTGCTG 540 GACTCCGACG GCTCCTTCTT CCTCTACAGC AAGCTCACCG TGGACAAGAG CAGGTGGCAG 600 CAGGGGAACG TCTTCTCATG CTCCGTGATG CATGAGGCTC TGCACAACCA CTACACGCAG 660 AAGAGCCTCT CCCTGTCTCC GGGTAAAGTA CCGATCCAGA AAGTTCAGGA CGACACCAAA 720 ACCTTAATTA AAACGATCGT TACGCGTATC AACGACATCA GTCACACCCA GTCGGTGAGC 780 TCTAAACAGA AAGTTACAGG CCTGGACTTC ATCCCGGGTC TGCACCCGAT CCTGACCTTG 840 TCCAAAATGG ACCAGACCCT GGCTGTATAC CAGCAGATCT TAACCTCCAT GCCGTCCCGT 900 AACGTTATCC AGATCTCTAA CGACCTCGAG AACCTTCGCG ACCTGCTGCA CGTGCTGGCA 960 TTCTCCAAAT CCTGCCACCT GCCATGGGCT TCAGGTCTTG AGACTCTGGA CTCTCTGGGC 1020

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GGGGTCCTGG AAGCATCCGG TTACAGCACC GAAGTTGTTG CTCTGTCCCG TCTGCAGGGT 1080
TCCCTTCAGG ACATGCTTTG GCAGCTGGAC CTGTCTCCGG GTTGTTAATG GATCC 1135

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1135 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GTATACCTGT TTTGAGTGTG TACAGGTGGA ACAGGTCGAG GCCTTGAGGA CCCCCCAGGA 60 AGTCAGAAGG AGAAGGGGG TTTTGGGTTC CTGTGGGAGT ACTAGAGGGC CTGGGGACTC 120 CAGTGTACGC ACCACCACCT GCACTCGGTG CTTCTGGGAC TCCAGTTCAA GTTGACCATG 180 CACCTGCCGC ACCTCCACGT ATTACGGTTC TGTTTCGGCG CCCTCCTCGT CATGTTGTCG 240 TGCATGGCAC ACCAGTCGCA GGAGTGGCAG GACGTGGTCC TGACCGACTT ACCGTTCCTC 300 ATGTTCACGT TCCAGAGGTT GTTTCGGGAG GGTCGGGGGT AGCTCTTTTG GTAGAGGTTT 360 CGGTTTCCCG TCGGGGCTCT TGGTGTCCAC ATGTGGGACG GGGGTAGGGC CCTACTCGAC 420 TGGTTCTTGG TCCAGTCGGA CTGGACGGAC CAGTTTCCGA AGATAGGGTC GCTGTAGCGG 480 CACCTCACCC TCTCGTTACC CGTCGGCCTC TTGTTGATGT TCTGGTGCGG AGGGCACGAC 540 CTGAGGCTGC CGAGGAAGAA GGAGATGTCG TTCGAGTGGC ACCTGTTCTC GTCCACCGTC 600 GTCCCCTTGC AGAAGAGTAC GAGGCACTAC GTACTCCGAG ACGTGTTGGT GATGTGCGTC 660 TTCTCGGAGA GGGACAGAGG CCCATTTCAT GGCTAGGTCT TTCAAGTCCT GCTGTGGTTT 720 TGGAATTAAT TTTGCTAGCA ATGCGCATAG TTGCTGTAGT CAGTGTGGGT CAGCCACTCG 780 AGATTTGTCT TTCAATGTCC GGACCTGAAG TAGGGCCCAG ACGTGGGCTA GGACTGGAAC 840 AGGTTTTACC TGGTCTGGGA CCGACATATG GTCGTCTAGA ATTGGAGGTA CGGCAGGGCA 900 TTGCAATAGG TCTAGAGATT GCTGGAGCTC TTGGAAGCGC TGGACGACGT GCACGACCGT 960 AAGAGGTTTA GGACGGTGGA CGGTACCCGA AGTCCAGAAC TCTGAGACCT GAGAGACCCG 1020 CCCCAGGACC TTCGTAGGCC AATGTCGTGG CTTCAACAAC GAGACAGGGC AGACGTCCCA 1080

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AGGGAAGTCC TGTACGAAA	C CGTCGACCTG	GACAGAGGCC	CAACAATTAC	CTAGG
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(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 374 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: protein
- (ix) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "Met (ATG) starts at -1"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
 - Met Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu 10
 - Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu
- Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser 40
- His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu 50 55
- Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr 65 70 80
- Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn 85
- Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro 100
- Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln 115 120 125
- Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val 130 135 140
- Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val 150 155 160

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Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro
165 170 175

Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr

Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val 195 200 205

Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu 210 215 220

Ser Pro Gly Lys Val Pro Ile Gln Lys Val Gln Asp Asp Thr Lys Thr 235 230 240

Leu Ile Lys Thr Ile Val Thr Arg Ile Asn Asp Ile Ser His Thr Gln 245 250 255

Ser Val Ser Ser Lys Gln Lys Val Thr Gly Leu Asp Phe Ile Pro Gly 260 265 270

Leu His Pro Ile Leu Thr Leu Ser Lys Met Asp Gln Thr Leu Ala Val 275 280 285

Tyr Gln Gln Ile Leu Thr Ser Met Pro Ser Arg Asn Val Ile Gln Ile 290 295 300

Ser Asn Asp Leu Glu Asn Leu Arg Asp Leu Leu His Val Leu Ala Phe 305 310 315 320

Ser Lys Ser Cys His Leu Pro Trp Ala Ser Gly Leu Glu Thr Leu Asp 325 330 335

Ser Leu Gly Gly Val Leu Glu Ala Ser Gly Tyr Ser Thr Glu Val Val 340 345 350

Ala Leu Ser Arg Leu Gln Gly Ser Leu Gln Asp Met Leu Trp Gln Leu 355 360 365

Asp Leu Ser Pro Gly Cys 370

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1135 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

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(ix) FEATURE:

(A) NAME/KEY: misc_feature

(B) LOCATION: 4

(D) OTHER INFORMATION: /note= "Met = ATG"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

(CATATGGACA	AAACTCACAC	ATGCCCACCG	TGCCCAGCTC	CGGAACTCGA	AGGTGGTCCG	60
•	ICAGTCTTCC	TCTTCCCCC	AAAACCCAAG	GACACCCTCA	TGATCTCCCG	GACCCCTGAG	120
(GTCACATGCG	TGGTGGTGGA	CGTGAGCCAC	GAAGACCCTG	AGGTCAAGTT	CAACTGGTAC	180
(GTGGACGGCG	TGGAGGTGCA	TAATGCCAAG	ACAAAGCCGC	GGGAGGAGCA	GTACAACAGC	240
2	ACGTACCGTG	TGGTCAGCGT	CCTCACCGTC	CTGCACCAGG	ACTGGCTGAA	TGGCAAAGCT	300
	PACGCATGCG	CGGTCTCCAA	CAAAGCCCTC	CCAGCCCCCA	TCGAGAAAAC	CATCTCCAAA	360
(GCCAAAGGGC	AGCCCCGAGA	ACCACAGGTG	TACACCCTGC	CCCCATCCCG	GGATGAGCTG	420
2	ACCAAGAACC	AGGTCAGCCT	GACCTGCCTG	GTCAAAGGCT	TCTATCCCAG	CGACATCGCC	480
(GTGGAGTGGG	AGAGCAATGG	GCAGCCGGAG	AACAACTACA	AGACCACGCC	TCCCGTGCTG	540
(GACTCCGACG	GCTCCTTCTT	CCTCTACAGC	AAGCTCACCG	TGGACAAGAG	CAGGTGGCAG	600
(CAGGGGAACG	TCTTCTCATG	CTCCGTGATG	CATGAGGCTC	TGCACAACCA	CTACACGCAG	660
2	AAGAGCCTCT	CCCTGTCTCC	GGGTAAAGTA	CCGATCCAGA	AAGTTCAGGA	CGACACCAAA	720
2	ACCTTAATTA	AAACGATCGT	TACGCGTATC	AACGACATCA	GTCACACCCA	GTCGGTGAGC	780
7	rctaaacaga	AAGTTACAGG	CCTGGACTTC	ATCCCGGGTC	TGCACCCGAT	CCTGACCTTG	840
7	rccaaaatgg	ACCAGACCCT	GGCTGTATAC	CAGCAGATCT	TAACCTCCAT	GCCGTCCCGT	900
2	AACGTTATCC	AGATCTCTAA	CGACCTCGAG	AACCTTCGCG	ACCTGCTGCA	CGTGCTGGCA	960
•	TTCTCCAAAT	CCTGCCACCT	GCCATGGGCT	TCAGGTCTTG	AGACTCTGGA	CTCTCTGGGC	1020
(GGGTCCTGG	AAGCATCCGG	TTACAGCACC	GAAGTTGTTG	CTCTGTCCCG	TCTGCAGGGT	1080
•	TCCCTTCAGG	ACATGCTTTG	GCAGCTGGAC	CTGTCTCCGG	GTTGTTAATG	GATCC	1135

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(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1135 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

60	TCCACCAGGC	GCCTTGAGCT	ACGGGTCGAG	TACGGGTGGC	TTTGAGTGTG	GTATACCTGT
120	CTGGGGACTC	ACTAGAGGGC	CTGTGGGAGT	TTTTGGGTTC	AGAAGGGGG	AGTCAGAAGG
180	GTTGACCATG	TCCAGTTCAA	CTTCTGGGAC	GCACTCGGTG	ACCACCACCT	CAGTGTACGC
240	CATGTTGTCG	CCCTCCTCGT	TGTTTCGGCG	ATTACGGTTC	ACCTCCACGT	CACCTGCCGC
300	ACCGTTTCGA	TGACCGACTT	GACGTGGTCC	GGAGTGGCAG	ACCAGTCGCA	TGCATGGCAC
360	GTAGAGGTTT	AGCTCTTTTG	GGTCGGGGGT	GTTTCGGGAG	GCCAGAGGTT	ATGCGTACGC
420	CCTACTCGAC	GGGGTAGGGC	ATGTGGGACG	TGGTGTCCAC	TCGGGGCTCT	CGGTTTCCCG
480	GCTGTAGCGG	AGATAGGGTC	CAGTTTCCGA	CTGGACGGAC	TCCAGTCGGA	TGGTTCTTGG
540	AGGGCACGAC	TCTGGTGCGG	TTGTTGATGT	CGTCGGCCTC	TCTCGTTACC	CACCTCACCC
600	GTCCACCGTC	ACCTGTTCTC	TTCGAGTGGC	GGAGATGTCG	CGAGGAAGAA	CTGAGGCTGC
660	GATGTGCGTC	ACGTGTTGGT	GTACTCCGAG	GAGGCACTAC	AGAAGAGTAC	GTCCCCTTGC
720	GCTGTGGTTT	TTCAAGTCCT	GGCTAGGTCT	CCCATTTCAT	GGGACAGAGG	TTCTCGGAGA
780	CAGCCACTCG	CAGTGTGGGT	TTGCTGTAGT	ATGCGCATAG	TTTGCTAGCA	TGGAATTAAT
840	GGACTGGAAC	ACGTGGGCTA	TAGGGCCCAG	GGACCTGAAG	TTCAATGTCC	AGATTTGTCT
900	CGGCAGGGCA	ATTGGAGGTA	GTCGTCTAGA	CCGACATATG	TGGTCTGGGA	AGGTTTTACC
960	GCACGACCGT	TGGACGACGT	TTGGAAGCGC	GCTGGAGCTC	TCTAGAGATT	TTGCAATAGG
1020	GAGAGACCCG	TCTGAGACCT	AGTCCAGAAC	CGGTACCCGA	GGACGGTGGA	AAGAGGTTTA
1080	AGACGTCCCA	GAGACAGGGC	CTTCAACAAC	AATGTCGTGG	TTCGTAGGCC	CCCCAGGACC
1135	CTAGG	CAACAATTAC	GACAGAGGCC	CGTCGACCTG	TGTACGAAAC	AGGGAAGTCC

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- (2) INFORMATION FOR SEQ ID NO:18:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 374 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: protein
 - (ix) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "Met (ATG) starts at -1"
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
 - Met Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Glu 10
 - Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu
 - Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser 40
 - His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu 50 55
 - Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr 65 70 75
 - Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn 85 90
 - Gly Lys Ala Tyr Ala Cys Ala Val Ser Asn Lys Ala Leu Pro Ala Pro 100 105 110
 - Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln 115 120
 - Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val 130 135
 - Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val 145 150 155 160
 - Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro 165 170 175

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Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr 180 185 190

Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val 195 200 205

Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu 210 215 220

Ser Pro Gly Lys Val Pro Ile Gln Lys Val Gln Asp Asp Thr Lys Thr 225 230 235 240

Leu Ile Lys Thr Ile Val Thr Arg Ile Asn Asp Ile Ser His Thr Gln 245 250 255

Ser Val Ser Ser Lys Gln Lys Val Thr Gly Leu Asp Phe Ile Pro Gly 260 265 270

Leu His Pro Ile Leu Thr Leu Ser Lys Met Asp Gln Thr Leu Ala Val 275 280 285

Tyr Gln Gln Ile Leu Thr Ser Met Pro Ser Arg Asn Val Ile Gln Ile 290 295 300

Ser Asn Asp Leu Glu Asn Leu Arg Asp Leu Leu His Val Leu Ala Phe 305 310 315 320

Ser Lys Ser Cys His Leu Pro Trp Ala Ser Gly Leu Glu Thr Leu Asp 325 330 335

Ser Leu Gly Gly Val Leu Glu Ala Ser Gly Tyr Ser Thr Glu Val Val 340 345 350

Ala Leu Ser Arg Leu Gln Gly Ser Leu Gln Asp Met Leu Trp Gln Leu 355 360 365

Asp Leu Ser Pro Gly Cys 370

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CLAIMS

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- 1. A protein having a formula selected from the group consisting of: $R_1 R_2$ and $R_1 L R_2$, wherein R_1 is a Fc protein or analog thereof, R_2 is an OB protein or analog thereof, and L is a linker.
- 2. The protein according to claim 1, where in the Fc, analog or derivative is selected from the group consisting of:
 - (a) the Fc amino acid sequences as set forth in SEQ. ID. NOS.: 9, 12, 15 and 18;
 - (b) the amino acid sequence of subpart (a) having a different amino acid substituted or deleted in one or more of the following positions (using the numbering according to SEQ. ID. NO. 9):
 - (i) one or more cysteine residues replace by an alanine or serine residue;
 - (ii) one or more tyrosine residues
 replaced by a phenylalanine residue;
 - (iii) the amino acid at position 5 replaced with an alanine;
 - (iv) the amino acid at position 20 replaced with glutamate;
- 25 (v) the amino acid at position 103 replaced with an alanine;
 - (vi) the amino acid at position 105 replaced with an alanine;
- (vii) the amino acid at position 107 replaced with an alanine;
 - (viii) the amino acids at positions 1,
 2, 3, 4, or 5 deleted;
 - (ix) one or more residues replaced or deleted to ablate the Fc receptor binding site;

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- one or more residues replaced or deleted to ablate the complement (Clq) binding site; and
 - (xi) a combination of subparts i-x;
- (c) the amino acid sequence of subparts (a)
 or (b) having a methionyl residue at the
 N-terminus;
- (d) the Fc protein, analog or derivative of any of subparts (a) through (c) comprised of a chemical moiety connected to the protein moiety;
- (e) a derivative of subpart (d) wherein said chemical moiety is a water soluble polymer moiety;
- (f) a derivative of subpart (e) wherein said water soluble polymer moiety is polyethylene glycol;
- (g) A derivative of subpart (e) wherein said water soluble polymer moiety is a polyamino acid moiety; and
- (h) a derivative of subpart (e) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.
- 3. The protein according to claim 1, wherein the OB protein, analog or derivative is selected from the group consisting of:
 - (a) the amino acid sequence 1-146 as set forth in SEQ. ID. NO. 3 or SEQ. ID. NO. 6;
 - (b) the amino acid sequence 1-146 as set forth in SEQ. ID. NO. 6 having a lysine residue at position 35 and an isoleucine residue at position 74;
 - (c) the amino acid sequence of subpart (b) having a different amino acid substituted in one or more of the following positions (using the numbering according to SEQ. ID. NO. 6): 4, 8, 32,

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- 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 112, 118, 136, 138, 142, and 145;
- (d) the amino acid sequence of subparts (a),(b) or (c) optionally lacking a glutaminyl residue at position 28;
- (e) the amino acid sequence of subparts (a),(b), (c), or (d) having a methionyl residue at the N-terminus.
- (f) a truncated OB protein analog selected from among: (using the numbering of SEQ. ID. NO. 6 having a lysine residue at position 35, and an isoleucine residue at position 74):
 - (i) amino acids 98-146
 - (ii) amino acids 1-32
 - (iii) amino acids 40-116
 - (iv) amino acids 1-99 and 112-146
 - (v) amino acids 1-99 and 112-146 having one or more of amino acids 100-111 sequentially placed between amino acids 99 and 112; and,
 - (vi) the truncated OB analog of subpart (f)(i) having one or more of amino acids 100, 102, 105, 106, 107, 108, 111, 112, 118, 136, 138, 142, and 145 substituted with another amino acid;
 - (vii) the truncated analog of subpart
 (f)(ii) having one or more of amino acids 4, 8 and
 32 substituted with another amino acid;
 - (viii) the truncated analog of subpart (f)(iii) having one or more of amino acids 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111 and 112 replaced with another amino acid:
 - (vix) the truncated analog of subpart
 (f)(iv) having one or more of amino acids 4, 8, 32,

- 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 112, 118, 136, 138, 142, and 145 replaced with another amino acid;
- (x) the truncated analog of subpart

 (f) (v) having one or more of amino acids 4, 8,32,

 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74,

 77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111,

 112, 118, 136, 138, 142, and 145 replaced with

 another amino acid;
- (xi) the truncated analog of any of subparts (f)(i)-(x) having an N-terminal methionyl residue;
 - (g) the OB protein or analog derivative of any of subparts (a) through (f) comprised of a chemical moiety connected to the protein moiety;
 - (h) a derivative of subpart (g) wherein said chemical moiety is a water soluble polymer moiety;
 - (i) a derivative of subpart (h) wherein said water soluble polymer moiety is polyethylene glycol;
 - (j) A derivative of subpart (h) wherein said water soluble polymer moiety is a polyamino acid moiety; and
- (k) a derivative of subpart (h) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.
- 4. The protein of claim 1 wherein the linker sequence is one or more amino acids selected from the group consisting of: Glycine, Asparagine, Serine, Threonine and Alanine.
 - 5. The protein of claim 1 wherein the linker is selected from the group consisting of:
- 35 (a) ala, ala, ala;
 - (b) ala, ala, ala, ala;

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(c)
                          ala, ala, ala, ala;
               (d)
                          gly, gly;
               (e)
                          gly, gly, gly;
                          gly, gly, gly, gly, gly;
               (f)
 5
                          gly, gly, gly, gly, gly, gly;
               (g)
               (h)
                          gly-pro-gly;
                          gly, gly, pro, gly, gly;
               (i)
                         chemical moiety; and
               (j)
                          any combination of subparts (a)
               (k)
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    through (j).
```

- A fusion protein comprising a Fc protein, analog or derivative thereof, fused to the N-terminus of an OB protein, analog or derivative 15 thereof.
- A nucleic acid sequence encoding for a 7. protein having the formula selected from the group consisting of: R_1 - R_2 and R_1 - L - R_2 , wherein R_1 is a Fc protein or analog thereof, R2 is an OB protein or analog 20 thereof, and L is a linker.
- The nucleic acid sequence according to 8. claim 7 encoding for a protein having a Fc, analog or derivative portion selected from the group consisting 25 of:
 - (a) the Fc amino acid sequences as set forth in SEQ. ID. NOS.: 9, 12, 15 and 18;
- (b) the amino acid sequence of subpart (a) having a different amino acid substituted or 30 deleted in one or more of the following positions (using the numbering according to SEQ. ID. NO. 9):
 - (i) one or more cysteine residues replace by an alanine or serine residue;

	(ii) one or more tyrosine residues
	replaced by a phenylalanine residue;
	(iii) the amino acid at position 5
	replaced with an alanine;
5	(iv) the amino acid at position 20
	replaced with glutamate;
4	(v) the amino acid at position 103
	replaced with an alanine;
	(vi) the amino acid at position 105
10	replaced with an alanine;
	(vii) the amino acid at position 107
	replaced with an alanine;
	(viii) the amino acids at positions 1
4.5	2, 3, 4, or 5 deleted;
15	(ix) one or more residues replaced
	or deleted to ablate the Fc receptor binding site;
	(x) one or more residues replaced
	or deleted to ablate the complement (Clq) binding
20	site; and
20	(xi) a combination of subparts i-x;
	(c) the amino acid sequence of subparts (a)
	or (b) having a methionyl residue at the
	N-terminus;
25	(d) the Fc protein, analog or derivative of
23	any of subparts (a) through (c) comprised of a
	chemical moiety connected to the protein moiety;
	(e) a derivative of subpart (d) wherein said chemical moiety is a water soluble polymer moiety;
	(f) a derivative of subpart (e) wherein said
30	water soluble polymer moiety is polyethylene
	glycol;
	(g) A derivative of subpart (e) wherein said
	water soluble polymer moiety is a polyamino acid
	moiety; and
	· · · · · · · · · · · · · · · · · · ·

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- a derivative of subpart (e) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.
- 5 The nucleic acid sequence according to 9. claim 7 encoding for a protein having an OB protein, analog or derivative portion selected from the group consisting of:
 - the amino acid sequence 1-146 as set (a) forth in SEQ. ID. NO. 3 or SEQ. ID. NO. 6;

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- the amino acid sequence 1-146 as set forth in SEQ. ID. NO. 6 having a lysine residue at position 35 and an isoleucine residue at position 74;
- 15 (c) the amino acid sequence of subpart (b) having a different amino acid substituted in one or more of the following positions (using the numbering according to SEQ. ID. NO. 6): 4, 8, 32, 33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77, 20 78, 89, 97, 100, 102, 105, 106, 107, 108, 111, 112, 118, 136, 138, 142, and 145;
 - (d) the amino acid sequence of subparts (a), (b) or (c) optionally lacking a glutaminyl residue at position 28;
- 25 the amino acid sequence of subparts (a), (b), (c), or (d) having a methionyl residue at the N terminus.
 - (f) a truncated OB protein analog selected from among: (using the numbering of SEQ. ID. NO. 6 having a lysine residue at position 35, and an isoleucine residue at position 74):
 - (i) amino acids 98-146
 - (ii) amino acids 1-32
 - (iii) amino acids 40-116
- (iv) amino acids 1-99 and 112-146

	(v) amino acids 1-99 and 112-146 having
	one or more of amino acids 100-111 sequentially
	placed between amino acids 99 and 112; and,
	(vi) the truncated OB analog of subpart
5	(f)(i) having one or more of amino acids 100, 102,
	105, 106, 107, 108, 111, 112, 118, 136, 138, 142,
	and 145 substituted with another amino acid;
	(vii) the truncated analog of subpart
	(f)(ii) having one or more of amino acids 4, 8 and
10	32 substituted with another amino acid;
	(viii) the truncated analog of subpart
	(f)(iii) having one or more of amino acids 50, 53,
	60, 64, 66, 67, 68, 71, 74, 77, 78, 89, 97, 100,
	102, 105, 106, 107, 108, 111 and 112 replaced with
15	another amino acid;
	(vix) the truncated analog of subpart
	(f)(iv) having one or more of amino acids 4, 8, 32,
	33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74, 77,
	78, 89, 97, 112, 118, 136, 138, 142, and 145
20	replaced with another amino acid;
	(x) the truncated analog of subpart
	(f)(v) having one or more of amino acids 4, 8,32,
	33, 35, 48, 50, 53, 60, 64, 66, 67, 68, 71, 74,
	77, 78, 89, 97, 100, 102, 105, 106, 107, 108, 111,
25	112, 118, 136, 138, 142, and 145 replaced with
	another amino acid;
	(xi) the truncated analog of any of
	subparts (f)(i)-(x) having an N-terminal methionyl
	residue;
30	
	(g) the OB protein or analog derivative of
	any of subparts (a) through (f) comprised of a
	chemical moiety connected to the protein moiety;
•	(h) a derivative of subpart (g) wherein said
35	chemical moiety is a water soluble polymer moiety;

	(i)	a d	erivativ	e of	subp	part	(h)	wherein	said
water	sol	uble	polymer	moie	ety i	s po	lyet	hylene	
glyco	1;								

- (j) A derivative of subpart (h) wherein said water soluble polymer moiety is a polyamino acid moiety; and
- (k) a derivative of subpart (h) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.

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10. The nucleic acid sequence of claim 7 encoding for a protein with a linker sequence of one or more amino acids selected from the group consisting of: Gly, Asn, Ser, Thr and Ala.

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- 11. The nucleic acid sequence of claim 7 encoding for a protein with a linker selected from the group consisting of:
 - (a) ala, ala, ala;
 - (b) ala, ala, ala;
 - (c) ala, ala, ala, ala;
 - (d) gly, gly;
 - (e) gly, gly, gly;
 - (f) gly, gly, gly, gly;
- 25 (g) gly, gly, gly, gly, gly, gly;
 - (h) gly-pro-gly;
 - (i) gly, gly, pro, gly, gly;
 - (j) a chemical moiety; and
 - (k) any combination of subparts (a)
- 30 through (j).
 - 12. A nucleic acid sequence encoding for a fusion protein having a Fc protein, analog or derivative thereof, fused to the N-terminus of an OB protein, analog or derivative thereof.

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- 13. A vector containing a nucleic acid sequence according to claims 7 or 12.
- 14. The vector of claim 13 wherein the vector is pAMG21 land the nucleic acid sequence according to claims 7 or 12.
 - 15. A prokaryotic or eukaryotic host cell containing the vector of claim 13.

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16. A process for producing a protein of claims 1 or 6 comprising the steps of culturing, under suitable conditions, the host cell of claim 15, and isolating the protein produced.

- 17. The process of claim 16 further comprising the step of purifying the protein produced.
- 18. A pharmaceutical composition comprising 20 an effective amount of a protein according to claims 1 or 6, in a pharmaceutically acceptable diluent, adjuvant or carrier.
- selected from the group consisting of excess weight, diabetes, high blood lipid level, arterial sclerosis, arterial plaque, the reduction or prevention of gall stones formation, insufficient lean tissue mass, insufficient sensitivity to insulin, and stroke, wherein the method consists of administering a therapeutically effective amount of the protein according to claims 1 or 6.

FIG 1A

σ	TCTAGATTTTGAGTTTTTAACTTTTTAGAAGAAGAATAACATATGGTACCGATCCAGAAAGT - 4	α
)	AGATCTAAACTCAAAAATCTTCCTCCTTATTGTATACCATGGCTAGGTCTTTCA M V P I Q K V	D I
Ċ	_	7
0 V	AGTCCTGCTGTGTTTTGGAATTTTTGCTAGCAATGCGCATAGTTGCTGTAGTCAGT Q D D T K T L I K T I V T R I N D I S H	φ 7 1 Ι
7		, ,
7 7	GTGGGTCAGCCAGAGGCGATTTGTCGCACAATGGCCAGACCTGAAGTAGGGCCCAGACGT T Q S V S A K Q R V T G L D F I P G L H	α α Η Ι
0	CCCGATCCTAAGCTTGTCCAAAATGGACCAGACCCTGGCTGTATACCAGCAGGTGTTAAC	0 7
184	_	47 J

	TTAATGGATCC	
ı	-	
488	GTCCCGTCTGCAGGGTTCCCTTCAGGACATCCTTCAGCAGCTGGACGTTTCTCCGGAATG	
) 1 !		
428	ACCGGAATCCCTGGACGGGTCCTGGAAGCATCCCTGTACAGCACCGAAGTTGTTGCTCT	
)))	309 -+	
8	GCTGCACCTGCAGATTCTCCAAATCCTGCTCCCTGCCGCAGACCTCAGGACTTTCAGAA	
)))	249 -+	
α	CTCCCTGCCGTCCCAGAACGTTCTTCAGATCGCTAACGACCTCGAGAACCTTCGCGACCT	

FIG 1E

FIG 2A

180 240 120 9 TGCGCATAGTTGCTGTAGTCAGTGTGGGTCAGCCACTCGAGATTTGTCGCACAATGTCCG CTGGACTTCATCCCGGGTCTGCACCCGATCCTGACCTTGTCCAAAATGGACCAGACCCTG **ACGCGTATCAACGACATCAGTCACCCAGTCGGTGAGCTCTAAACAGCGTGTTACAGGC** GACCTGAAGTAGGGCCCAGACGTGGGCTAGGACTGGAACAGGTTTTACCTGGTCTGGGAC GCTGTATACCAGCAGATCTTAACCTCCATGCCGTCCCGTAACGTTCTTCAGATCTCAAAC CGACATATGGTCGTCTAGAATTGGAGGTACGGCAGGGCATTGCAAGAAGTCTAGAGATTG Z S Ø E ĸ 区 Ø Ø Σ H Н \Box 봈 × > ഗ W H Z 又 S Ы 召 \vdash > H S Ø Ω Ч Д П Ø Н Σ Ø Д ۲ Ŋ > H 二 EH ᅜ ഗ Ц Ц Ø Ü Д \vdash Д Ø Д Z α > \succ Ω Σ 吖 > Ы E K 121

300 420 360 GACCTCGAGAACCTTCGCGACCTGCTGCTGCTGGCATTCTCCAAATCCTGCCACCTG CTGGAGCTCTTGGAAGCGCTGGACGACGTGCACGACCGTAAGAGGTTTTAGGACGGTGGAC CCATGGGCTTCAGGTCTTGAGACTCTGGACTCTCTGGGCGGGGTCCTGGAAGCATCCGGT GGTACCCGAAGTCCAGAACTCTGAGACCTGAGAGCCCCGCCCCAGGACCTTCGTAGGCCA TACAGCACCGAAGTTGTTGCTCTGTCCCGTCTGCAGGGTTCCCTTCAGGACATGCTTTGG ATGTCGTGGCTTCAACAACGAGACAGGGCAGACGTCCCAAGGGAAGTCCTGTACGAAACC 3 Ц Σ Д × Ø ഗ Ц Ö ഗ 压 454 Ö Þ O Ø GTCGACCTGGACAGGCCCAACAATTACCTAGG CAGCTGGACCTGTCTCCGGGTTGTTAATGGATCC ഗ 耳 Ω ᄓ Ц Ŋ П E Д 口 ΛΛ ĸ П Ы Ö 口 Z Ŋ 闰 Ħ Ы 3 S Д Д 241 301 361 421

FIG 2E

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FIG 3A-1

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240 120 180 9 GAGGACCCCCCTGGCAGTCAGAAGGAGAGGGGGGTTTTTGGGTTCCTGTGGGAGTACTAG AAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAG GTATACCTTGGGTTTAGAACACTGTTTTGAGTGTGTACGGGTGGCACGGGTCGTGGACTT TCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTC AGGGCCTGGGGACTCCAGTGTACGCACCACCTGCACTCGGTGCTTCTGGGACTCCAG CTCCTGGGGGGACCGTCAGTCTTCCTCTCCCCCAAAACCCCAAGGACACCCTCATGATC TTCAAGTTGACCATGCACCTGCCGCACCTCCACGTATTACGGTTCTGTTTCGGCGCCCTC CATATGGAACCCAAATCTTGTGACAAAACTCACACATGCCCACCGTGCCCAGCACCTGAA Ш 山 Σ $\Omega_{\mathbf{i}}$ ы Д Ø О ۲ Д Е Д **[11**] \mathbf{C} × × 耳 Д Д Д S > × Z O Ω Д \vdash > 二 Д > . ⊟ ſμ > Ц 姳 U Ω ſτι > H \mathcal{O} > ഗ S 回 Д × U рų Д Ü 团 Е 니 ø Σ' ᆸ Ŋ 61 121 181

FIG 3A-2

FIG 3B-1

ACGCCTCCCGTGCTGGACTCCGACGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGAC +++++++	u Q	AAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCAC		H	AACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAAGTACCGATCCAGAAAGTT+++++++-	· >	CAGGACGACACCAAAACCTTAAAACGATCGTTACGCGTATCAACGACATCAGTCAC		1
CGT(GCA(DGSFFLYSKLTV	TCT(AGA(A	GAAJ CTT	SLSPGKVPIQKV	CAG	3TC2	H S I C N I S II N I S I
CAC GTG	E	GGC	CCG		CCA GGT	Q	CAT	GTA(-
GCT ++	ъ	TGA	ACT	团	GAT + CTA	Н	CGA	GCT	ב
SCAP	幺	ACD:	\CGT	耳	PACC 	Дŧ	CAA	GTT	Z
ACAC	ഗ	rga1	ACTP	SCSVMHE	AAGT ITCA	>	FTAT	ATA	-
TCT7	≯	CCG.	3GC7	>	3TA/ +-	×	2GCC	308(Ω
TCC.	니	3CT(CGA(Ŋ	C 3CC(Ö	rta(AAT(E-
TCT	ĮΤΙ	CAT	3TA(Ü	CTC	Д	TCG	AGC.	\triangleright
CCT.	Įτι	rc T	AGA(Ŋ	IGT(-+-	Ŋ	CGA	, 3CT2	I -
GGA(S	ICT	4GA.	> ₽	799g	Ы	AAAC	LT.I	E-
ACG(O	ACG	IGC/	N	ICT(AGA(L S	rta.	4AT	×
2000 3000	О	GGA	CCT		3000	IJ	raa:	ATTZ	 -
ACTO	ß	AGG(TCC	Ŋ	AGA(T	Ŋ	CCT	3GA1	
TGG	L D	AGC.	TCG'	Ø	AGA.	×	AAA(TTT(E-
TGC ACG		GGC	SSS	Ø	 	Q F	CCA	GGT	×
000 +	>	GGT	CCA	3	CACTACACGCAGA + GTGATGTGCGTCT	H	ACA +	TGT	E-
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ACGCCTCCGTGCTGGACT + TGCGGAGGCCACGACCTGA	Ω,	AAGAGCAGGTGGCAGG	TCT	S	AACC	H	CAGGACGACACCAAAACCT	TCC	C
541 -	₽	4 LO2		×	A 661 -	4	721		ر

FIG 3B-2

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1020 1080 840 900 960 **ACCCAGTCGGTGAGCTCTAAACAGAAAGTTACAGGCCTGGACTTCATCCCGGGTCTGCAC** TGGGTCAGCCACTCGAGATTTGTCTTTCAATGTCCGGACCTGAAGTAGGGCCCAGACGTG GGCTAGGACTGGAACAGGTTTTTACCTGGTCTGGGACCGACATATGGTCGTCTAGAATTGG AGGTACGGCAGGGCATTGCAATAGGTCTAGAGATTGCTGGAGCTCTTGGAAGCGCTGGAC CCGATCCTGACCTTGTCCAAAATGGACCAGACCCTGGCTGTATACCAGCAGATCTTAACC TCCATGCCGTCCCGTAACGTTATCCAGATCTCTAACGACCTCGAGAACCTTCGCGACCTG CTGCACGTGCTGGCATTCTCCAAATCCTGCCACCTGCCATGGGCTTCAGGTCTTGAGACT CTGGACTCTCTGGGGGGTCCTGGAAGCATCCGGTTACAGCACCGAAGTTGTTGCTCTG GACGTGCACGACCGTAAGAGGTTTAGGACGGTGGACGGTACCCGAAGTCCAGAACTCTGA GACCTGAGAGCCCCCCCAGGACCTTCGTAGGCCAATGTCGTGGCTTCAACAACGAGAC 工 ⊱ П Д 山 Ø ග Н Ø O Z Ø ß [1] ഠ Ы > Д H 3 ഗ Ø Ω Д Ħ Н O Z Ы O ഗ E-1 H 二 Ŋ Ø Н U ď 노 Ø Д ഗ ſij Ø Σ × Ц × X Ŋ > Z ß S Ö ø ø S Ç Ŋ Н 口 ഗ S 田 Н Ø Σ О П Н Е ഗ 901 961 841 1021

FIG 30

1140 t TCCCGTCTGCAGGGTTCCCTTCAGGACATGCTTTGGCAGCTGGACCTGTCTCCGGGTTGT AGGGCAGACGTCCCAAGGGAAGTCCTGTACGAAACCGTCGACCTGGACAGAGGCCCAACA \mathcal{O} Ŋ Д ß Н Д 긔 Ø 3 Ы \mathbf{z} О Ø Ц ഗ ----+ 1150 ATTACCTAGG Ü Ø TAATGGATCC 니 民 S 1081

FIG 4A-1

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1	[II]	CTCCTGGGGGGTCCTTCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGATC	GAGGACCCCCCAGGAAGTCAGAAGGAGAAGGGGGGTTTTTGGGTTCCTGTGGGAGTACTAG	VFLFPPKPKDTLMI-	TCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTC		^	AAGTTCAACTGGTACGTGGACGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAG	TTCAAGTTGACCATGCACCTGCCGCACCTCCACGTATTACGGTTCTGTTTCGGCGCCCTC
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AGG	Д	CAC +	GTG	E-I	AGA +	TCT	Д	AAA	TTT
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AGG	വ	ACC -+-	${ m TGG}$	Д	GAG -+-	CTC	Ŋ	TGC	ACG
TAC	Ö	AAA 	TII	×	CGT	GCA	>	TAA	ATT
GTG	E		999	Д	GGA	CCT	Д	GCA	CGT
AGT	田	CCC +	GGG	Д	GGT +	CCA	>	GGT	CCA
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TTT.	봈	ICC.	AGG.	М	TGA(ACT	ഠ	3TA(CAT
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420		61
1	NGKEYKCKVSNKALPAPI	
360	CTGAATGGCAAGGAGTÀCAAGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCCATCGAG +++++	301
	TCGTGCATGGCAGTCGCAGGACGTGGTCCTGACC	- 1 #
300	GAGCAGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGG	241

FIG 4A-2

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E	ACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGAC ++++	Д	AAGAGCAGGTGGCAGCAGGGGAACGTCȚTCTCATGCTCCGTGATGCATGAGGCTCTGCAC +++++++	H	AACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAAGTACCGATCCAGAAGTT ++++++	>			ıı
×	GTG	>	CTG	J	AAA T'T'T	×	AGT 	TCA	S
×	ACC TGG	H	GCT CGA	A	CAG GTC	Ø	ATC	I'AG	Н
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田	ACTC	W	1666 1-1	Ŋ	GAG ++	S	CTI +	GAA	H
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æ	GCT 		GCA CGT	8	GCA CGT	Ø	CAA 	GTT	×
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ഗ	ACGCCTCCCGTGCTGGAC +TGCGGGCACGACCTG	ф	GAG	Ø	CCA	Ħ	3GA(GTCCTGCTGTGGTTTTGGAATTAATTTTGCTAGCAATGCGCATAGTTGCTGTAGTCAGTG	А
വ		댐	AA. 	×		z	CA(GT(Ø
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ŀ	LDSLGGVLEASGYSTEVVAL	
0 8 0 7	GACCTGAGAGCCCCCCAGGACCTTCGTAGGCCAATGTCGTGGCTTCAACAACGAGAC	
0	CTGGACTCTCTGGGGGCGGGTCCTGGAAGCATCCGGTTACAGCACCGAAGTTGTTGCTCTG	1021
1	LHVLAFSKSCHLPWASGLET	
0 7 0 7	GACG	
, ,	CTGCACGTGCTGGCATTCTCCAAATCCTGCCACCTGCCATGGGCTTCAGGTCTTGAGACT	o
t	SMPSRNVIQISNDLENLRDL	
0 9 6	AGGTACGGCAGGGCATTG	1
(TCCATGCCGTCCCGTAACGTTATCCAGATCTCTAACGACCTCGAGAACCTTCGCGACCTG	σ
1	PILTLSKMDQTLAVYQQILT	
)))	GGCTAGGACTGGAACA	,
(5	α
ı	TQSVSSKQKVTGLDF.IPGLH	
α4 Ο	TGGGTCAGCTCGAGATTTGTCTTTCAATG	-
(GGTCT	ι-

FIG 4B-2

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FIG 4C

TCCCGTCTGCAGGGTTCCCCTTCAGGACATGCTTTGGCAGCTGGACCTGTCTCCGGGTTGT AGGGCAGACGTCCCAAGGGGAAGTCCTGTACGAACCGTCGACCTGGACAGAGGCCCAACA C Ö Д ß Ц Ω Ц Ø 3 ᆸ Σ Ω Ø Ц ഗ 1150 Ŋ Ø ATTACCTAGG TAATGGATCC 니 ፈ ഗ 1081

FIG 5A-1

FIG 5A-2

540	•	
, ,	GTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTG	481
ı	TKNQVSLTCLVKGFYPSDIA	
480	TGGTTCTTGGTCCAGTCGGACTGGACGGACCAGTTTCCGAAGATAGGGTCGCTGTAGCGG	1, 1
(ACCAAG	421
1	AKGQPREPQ·VYTLPPSRDEL	
420	GCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAGCTG 1++++++++-	361
ı	Y.K.C.K.V.S.N.K.A.L.P.A.P.I.E.K.T.I.S.K	
360	TACAAGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCCATCGAGAAACCATCTCCAAA 1++++++	301
į	T Y R V V S V L T V L H Q D W L N G K E	
300	ACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGAG	241

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>	GTG TTG	M	CAC	GTG	E	CAC	3TG	Ę	3GT	CA
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Z	CTC	H	GAG	CTC	ы	ATC	TAG	н	GAC	CTC
Z	AAG 	X	CAI	GTA	田	0 0 0	099	Д	AAC	TTG
囙	AGC +	W.	ATG	TAC	Σi	GTA	CAT	>	ATC	TAG
G Q P E N N Y K T T P	TAC	≯	GTG	CAC	>	AAA	TTT	×	CGT	GCA
α	CTC GAG	LYSKLTVDKSRW	TCC	AGG	SVMHEALHNHYTQ	GGT	CCA	PGKVPIQKVQDDT	ACG	TGC
ტ	TTC -+- AAG	[z ₁	TGC	ACG	U	SCG F) 1 1 1 1 1	Д	GTTS	CAA
	FTC.	ĮŦ	rca	AGT.	ഗ	rct	4GA(ß	4TC	rag(
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回	GACTCCGACGGCTCCTTC +	S	CAGGGGAACGTCTTCTC	TCC	r O	AAGAGCCTCTCCCT	TCT		CCT	GGA
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1	G V L E A S G Y S T E V V A L S R L Q G	
1080	GOOGLOCI GGRAGORI COGGI I ROBOROCGARGITTGOTOTOTOTOTOTOTOTOTOTOTOTOGGGT	1021
1	FSKSCHLPWASGLETLDSLG	
1020	TTCTCCAAATCCTGCCACCTGCCATGGGCTTCAGGTCTTGAGACTCTGGACTCTCGGGC++++++ AAGAGGTTTAGGACGGTGCGTACCCGAAGTCCAGAACTCTGAGACCTGAGAGACCCG	961
ı	N V I Q I S N D L E N L R D L L H V L A	
096	AACGTTATCCAGATCTCTAACGACCTCGAGAACCTTCGCGACCTGCTGCACGTGCTGGCA 1++++++++ TTGCAATAGGTCTAGAGGTTTGCTGGAGCTCTTTGGAAGCGCTGGACGACGTGCACGTCGCGT	901
1	S K M D Q T L A V Y Q Q I L T S M P S R	
006	TCCAAAATGGACCAGACCCTGGCTGTATACCAGCAGATCTTAACCTCCATGCCGTCCCGT 1++++++++	841
1	SKOKVTGLDFIPGLHPILTL	
840	1CTRAACAGAAAGTTACAGGCCTGGACTTCATCCCGGGTCTGCACCCGATCCTGACCTTG 1+++++++ AGATTTGTCTTTCAATGTCGGACCTGAAGTAGGGCCCAGACGTGGGCTAGGACTGGAAC	781

FIG 5C

1135 TCCCTTCAGGACATGCTTTGGCAGCTGGACCTGTCTCCGGGTTGTTAATGGATCC AGGGAAGTCCTGTACGAACCGTCGACCTGGACAGAGGCCCCAACAATTACCTAGG U Ü Д ഗ Ц Ò 니 Ø 3 ᆸ Σ Ω Ø 니 ഗ 1081

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FIG 6A-1

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120 180 240 9 CATATGGACAAAACTCACACATGCCCACCGTGCCCAGCTĈCGGAACTCGAAGGTGGTCCG GTATACCTGTTTTGAGTGTGTACGGGTGGCACGGGTCGAGGCCTTGAGCTTCCACCAGGC TCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAG AGTCAGAAGGAGAAGGGGGTTTTGGGTTCCTGTGGGAGTACTAGAGGGCCTGGGGACTC CAGTGTACGCACCACCTGCACTCGGTGCTTCTGGGACTCCAGTTCAAGTTGACCATG CACCTGCCGCACCTCCACGTATTACGGTTCTGTTTCGGCGCCCTCCTCGTCATGTTGTCG GTCACATGCGTGGTGGACGTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTAC GTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAAGCCGCGGGAGGAGCAGTACAACAGC Д 团 ပ 口 以 Ŀ ഗ ᆸ 又 > 回 Н Σ ſτ] Д ᆸ Дı Ø H Д Ω Д O 回 봈 Д 耳 Д Д വ ပ 노 D V Д H > Д 耳 > \vdash U Д E Σ > ഗ 61 121 181

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5.40	GTGG	•
i	TKNQVSLTCLVKGFYPSDIA	
) 1	TGGTTCTTGGTCCAGTCG	T 7 T
480	·	``
1	AKGOPREPOVYTLPPSRDEL	
) 1	CGGTTTCCCGTCGGGGCTCTTGGTGTCCACATGTGGGGACGGGGGTAGGGCCCTACTCGAC	361
420	GCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAGCTG	ć
ı	YACAVSNKALPAPIEKTISK	
)))	ATGCGTACGCGCCAGAGGTTGTTTCGGGAGGGTCGGGGGGTAGCTCTTTTGGTAGAGGTTT	301
098	TACGCATGCGCGGTCTCCAACAAGCCCTCCCAGCCCCCATCGAGAAAACCATCTCCAAA	(
ı	TYRVVSVLTVLHQDWLNGKA	
	TGCATGGCACACCAGTCGCAGGAGTGGCAGGACGTGGTCCTGACCGACTTACCGTTTCGA	T # 7
300	ACGTACCGTGTGGTCAGCGTCCTCACCGTCCTTGCACCAGGACTGGCTGAATGGCAAAGCT	(

FIG 6A-2

FIG 6B-1

1	009	1	099	1	720	1	700	0	1
VEWESNGQPENNYKTTPPVL	GACTCCGACGCTCCTTCTTCCTCTACAGCAGCTCACCGTGGACAAGAGCAGGTGGCAG 541+++++++	DSDGSFFLYSKLTVDKSRWQ	CAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACACGCAG 601++++++++-	QGNVFSCSVMHEALHNHYTQ	AAGAGCCTCTCCCTGTCTCCGGGTAAGTACCGATCCAGAAAGTTCAGGACGACGACCAAA 661++++++++-	KSLSPGKVPIQKVQDDTK	ACCTTAATTAAAACGATCGTTACGCGTATCAACGACATCAGTCACGCCAGTCGGTGAGC	TGGAATTAATTTGCTAGCAATGCGCATAGTTGCTGTAGTCAGTGTGGGTCAGCCACTCG	TLIKTIVTRINDISH [*] TQSVS
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840) 1	I	006))	1	096	1	(, , , , ,	ı		1080	ı
TCTAAACAGAAAGTTACAGGCCTGGACTTCATCCCGGGTCTGCACCCGATCCTGACCTTG		S K Q K V T G L D F I P G L H P I L T L	TCCAAAATGGACCAGACCC,TGGCTGTATACCAGCAGATCTTAACCTCCATGCCGTCCCGT	CAGGGCA	SKMDQTLAVYQQILTSMPSR.	AACGTTATCCAGATCTCTAACGACCTCGAGAACCTTCGCGACCTGCTGCACGTGCTGGCA	NVIQISNDLENLRDLLHVLA	TTCTCCAAATCCTGCCACCTGCCATGGGCTTCAGGTCTTGAGACTCTGGACTCTGGGGC	AAGAGGTTTAGGACGGTGGACGGTACCCGAAGTCCAGAACTCTGAGACCTGAGAGCCCG	FSKSCHLPWASGLETLDSLG.	GGGGTCCTGGAAGCATCCGGTTACAGCACCGAAGTTGTTGCTCTGTCCCGTCTGCAGGGT	CCCCAGGACCTTCGTAGGCCAATGTCGTGGCTTCAACAACGAGACAGGGCAGACGTCCCA	GVLEASGYSTEVVALSRLQG
78			841			901		961	Ś		1001) 	

FIG 6C

1135 TCCCTTCAGGACATGCTTTGGCAGCTGGACCTGTCTCCGGGTTGTTAATGGATCC AGGGAAGTCCTGTACGAACCGTCGACCTGGACAGAGGCCCCAACAATTACCTAGG C Ü Ω, Ŋ П Ω 口 Ø Z П Σ Ω Ø П S

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A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/62 C07K14/575 A61K38/17

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

 $\begin{array}{ll} \mbox{Minimum documentation searched (classification system followed by classification symbols)} \\ IPC~6~~C12N~~C07K~~A61K \end{array}$

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUM	ENTS CONSIDERED TO BE RELEVANT	
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X Further documents are listed in the continuation of box C.	Patent family members are listed in annex.
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Date of the actual completion of the international search	Date of mailing of the international search report
3 April 1998	3 0. 04. 98
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk	Authorized officer
Tel. (+31-70) 340-2040, Tx. 31 651 epo ni, Fax: (+31-70) 340-3016	Hermann, R

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Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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